

Osteonecrosis of the Jaw—a Bone Site-Specific Effect of Bisphosphonates

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Abstract A known complication that can occur in patients using bisphosphonates (BPs) is osteonecrosis of the jaw (ONJ). ONJ features bone exposure that may be associated with severe pain, swelling, local infection, and pathological fracture of the jaw. Current literature indicates that a complex combination of factors is necessary to induce ONJ. Several hypotheses about the pathophysiology of ONJ were previously reported. Here, we review these hypotheses and introduce new ideas and suggestions on this topic, focusing on bone site-specific cells, and the effect that BPs and other anti-resorptive drugs have on those cells. Gaining more insight into bone site-specific effects may help to better understand the pathogenesis ONJ, and contribute to the development of new bone site-specific anti-resorptive drugs.

Keywords Bisphosphonates · Osteonecrosis · Jaw · ONJ · Osteoclast · Microenvironment · Bone marrow

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Introduction

Bisphosphonates (BPs) have been used for decades to treat bone-degrading diseases such as osteoporosis and bone metastasis. BPs increase bone density and reduce fracture risk by inhibiting bone resorption by osteoclasts. First-generation BPs do this by causing osteoclast apoptosis whereas the newer and more potent BPs containing nitrogen, such as alendronate (ALN) and zoledronic acid (ZA), also inhibit osteoclast activity [1–4]. By virtue of their high affinity for calcium and therefore rapid localization to the bone, BPs primarily act on osteoclasts because they are taken up by those cells during bone resorption [5]. A rare though severe side effect of BP use is osteonecrosis of the jaw (ONJ).

Osteonecrosis of the Jaw

ONJ was described as the first long-term complication of BP use [6–8]. With an incidence of up to 5–20 % after intravenous administration, the risk is much higher than after oral use, which has an incidence of around 0.04 % [9–11]. In 64 % of the cases, ONJ is preceded by a dental extraction or other oral trauma [12], and periodontitis often (84 %) accompanies ONJ [13]. In 2007, BP-related ONJ was defined as an area of exposed bone that did not heal within 8 weeks and was not the result of radiotherapy in the craniofacial region [14]. Recently, the International Task Force on Osteonecrosis of the Jaw updated the definition of ONJ from being a result of exposure to BPs to exposure to an anti-resorptive agent, indicating that BPs are not the sole initiators of ONJ [15•].

Clinically, apart from exposed or necrotic bone, ONJ can be recognized but not diagnosed by symptoms and radiographic observations such as pain, osteolysis, bone sequestration, and infection or soft tissue inflammation [15•, 16]. The presence

and severity of these symptoms are used to determine the stage of the disease according to the staging system first introduced by Ruggiero and co-workers [17] and later adopted by the American Association of Oral and Maxillofacial Surgeons (AAOMS) [16]. Several of the mentioned clinical manifestations were shown to be inducible in animal models which are partly listed in Table 1 and which will be discussed below.

Several hypotheses about the pathophysiology of ONJ were previously reviewed [11, 15, 34, 35]. Briefly, BPs may inhibit cells other than osteoclasts such as epithelial cells. For instance, a toxic effect on soft tissue could secondarily lead to bone exposure, and a negative effect on endothelial cells and/or angiogenesis may cause avascular necrosis [36, 37]. Microbial infections may lead to bone degradation directly, which may be worsened by a decreasing pH due to the inflammation [35, 38]. A reduced pH may also lead to release of BPs from the bone [39] and therefore be toxic to other cells. Furthermore, suppression of bone turnover, and the inability to heal microdamage, may lead to the accumulation of necrotic tissue [40, 41]. Finally, more recent studies show that single

nucleotide polymorphisms (SNPs) in several genes may lead to a genetic predisposition to ONJ [42, 43]. Altogether, a complex combination of factors seems to be necessary to induce ONJ, and more research is required to distinguish the roles of osteoclasts and bone loss in the pathogenic mechanism.

Animal Models

To gain more insight into the pathophysiology of osteonecrosis of the jaw, animal disease models were developed. Since an association between the RANKL inhibitor denosumab and ONJ was only first shown in 2010 [44, 45], the majority of animal models use BPs to induce ONJ. Since BPs alone did not usually lead to apparent signs of ONJ [46, 47], other inducing factors such as tooth extraction, immunosuppressive drugs, or periodontal disease were applied. Those combinations of treatments had variable outcomes on the manifestations of ONJ such as exposed bone, necrotic bone, bony sequestrum, and inflammation. An extensive overview

Table 1 Overview of animal models of ONJ

1 st author year	Species	Treatment	Frequency	Comorbidity	Exposed bone	Necrotic bone	Sequestrum	Inflammation	Ref
Abtahi 2013	Rat	sc ALN 200 ug/kg	1/day 3 weeks	DEX, extraction	10/10	10/10	6/10	10/10	[18]
Aghaloo 2011	Rat	ip ZA 66 ug/kg	3/week 15 weeks	Periodontal disease	4/19	9/19, area: ±60 %	6/19	Only with ligature	[19]
Aghaloo 2014	Mouse	ip OPG-Fc 10 mg/kg	1/week 12 weeks	Periapical disease	3/10	10/10, area: 17.7 %	ns	At sites of periapical disease	[20]
Aguirre 2012	Rat	iv ZA 80 ug/kg	1/month 18 weeks	Periodontal disease	Yes	Yes, apoptotic osteocytes	ns	Bacterial colonization	[21]
Allen 2008	Dog	Oral ALN 1 mg/kg	1/day 3 years	–	No	4/12	ns	ns	[22]
Allen 2013	Dog	iv ZA 60 ug/kg	1/2 weeks 9 months	DEX, extraction	No	No	No	Yes	[23]
Bi 2010	Mouse	ip ZA 125 ug/kg	2/week 15 weeks	DEX, docetaxel, extraction	10/10	10/10	Yes	10/10	[24]
de Molon 2014	Mouse	ip ZA ^a 200 ug/kg	3/week 12 weeks	Peri-radicular infection (natural)	8/38	10/38 hemimaxillae, area: 16 %	ns	Infection necessary for necrosis	[25••]
Hokugo 2013	Rat	iv ZA 70 ug/kg	1/month 2 months	VitD deficiency, extraction	ns	±55 % viable osteocytes	Yes	Yes	[26]
Howie 2015	Rat	iv ZA 80 ug/kg	1/week 13 weeks	Extraction (twice)	Yes	26.7 % empty lacunae	Yes	No bacterial colonization	[27]
Huja 2011	Dog	iv ZA 100 ug/kg	1/month 4 months	Extraction, mini-implant	No	Osteocyte death in maxilla, area: <1 %	ns	ns	[28]
Kikuri 2010	Mouse	iv ZA 125 ug/kg	2/week 8 weeks	DEX, extraction	Yes	Yes, area: ±5 %	Yes	Bacterial colonization and inflammation	[29]
Kuroshima 2012	Mouse	sc ZA 50 ug/kg	2/week 13 months	Excision palatal mucosa	ns	Empty lacunae	ns	PMN influx, no bacterial colonization	[30]
Kuroshima 2013	Mouse	sc ZA 50 ug/kg	2/week 7 weeks	MEL, extraction	13/16 sockets	Area: ±10 %	ns	PMNs but not macrophages	[31]
Pautke 2012	Mini-pig	iv ZA 50 ug/kg	1/week 16 weeks	Extraction	5/5	29/30 extraction sites	Yes	Bacterial colonization	[32]
Takaoka 2015	Rat	iv ZA 35 ug/kg	1/2 weeks 29 weeks	Diabetes, extraction	6/6	Empty lacunae	Yes	Bacterial colonization	[33]

When more than one treatment or doses are described in one paper, only the most relevant treatment for these signs of ONJ was mentioned in the table. Except in “frequency,” ## indicates the number of mice that show signs of the indicated pathology per total number of mice, unless indicated otherwise. ALN alendronate, DEX dexamethasone, MEL melphalan, ns not specified, PMN polymorphonuclear cell, VitD vitamin D, ZA zoledronic acid

^a Similar results with RANK-Fc and OPG-Fc

of ONJ manifestations is supplied in Table 1. Interestingly, the comorbidities of pro-inflammatory [19, 21, 25••] as well as anti-inflammatory [18, 23, 24, 29, 31] conditions induced symptoms such as exposed and necrotic bone.

Most models focus on the jaw alone, since osteonecrosis only occasionally occurs in other bones. Thus, it is appropriate to address the micro-environmental differences between the jaw and other bones, in both physiological and pathological circumstances.

Jaw Versus Other Bones

Long bone is produced via a cartilage intermediary, i.e., endochondral bone formation, whereas the jaw is primarily of intramembranous origin. The human mandible [48] as well as the murine calvaria [49] were shown to contain more collagen than long bones. Calcium content is also higher in the human mandible [48], and so it may adsorb more BPs than the long bones [50]. Besides the differential composition for various bones in the skeleton, the cells degrading and forming the osseous tissue, as well as the bone marrow composition, were shown to be bone site-specific.

Osteoclasts

Different bones have osteoclasts with distinct characteristics (reviewed in [51, 52]). Mechanistically, resorption by long-bone osteoclasts is mainly carried out by cathepsins whereas calvaria osteoclasts may also utilize matrix metalloproteinases (MMPs) [53]. Interestingly, MMP-9 was shown to play an important role in osteoclast-mediated stimulation of angiogenesis [54], indicating that it would be of interest to compare this interaction in different bones. Zenger et al. [55] showed higher expression of cathepsin K as well as tartrate-resistant acid phosphatase (TRACP) in long bone compared to calvaria osteoclasts. Long-bone osteoclasts also use different ion exchangers to maintain intracellular pH [56]. In vitro assays show different rates of osteoclastogenesis and RANKL/OPG ratios [57] as well as different osteoclastogenic potential [58] using mandibular and long-bone marrow. Furthermore, in response to BPs, jaw osteoclasts were less apoptotic compared to long-bone osteoclasts, indicating that the former may be less sensitive to BPs [59•]. Interestingly, in a mouse model for ONJ, RANKL inhibitors were more potent than ZA in reducing osteoclast numbers in the jaw [25••].

Osteoblasts

Similar to osteoclasts, bone-forming osteoblasts and their precursors in the bone marrow also differ among bone sites. Rat

mandibular-bone-marrow stromal cells were shown to have a higher osteogenic potential than those from tibia in vitro [60, 61]. Also in vitro, human jaw-derived bone marrow stromal cells demonstrated more calcium accumulation than iliac crest-derived cells whereas the latter induced more bone formation in mice under osteogenic conditions [62]. Matsubara and colleagues on the other hand showed similar osteogenic potential of human alveolar and iliac bone marrow stroma, but showed lower adipogenic and chondrogenic potential of alveolar BMSCs [63].

Next to differences in osteoblastogenic potential of stromal cells, osteoblast signaling towards other cells was shown to be bone site-dependent. Murine calvaria osteoblasts more potently induced osteoclastogenesis than those from long bone [64]. Human maxillary BMSCs were more potent inducers of angiogenesis than those derived from iliac crest [65]. On the other hand, disrupted *Vhl* gene expression in osteoblasts, and the resulting Hif α overexpression increased long-bone volume, but did not affect calvaria bones, indicating that osteoblast-stimulated angiogenesis plays an important role mainly in endochondral bone formation [66].

Even though BPs primarily affect osteoclasts, inhibiting effects on osteoblast viability and proliferation were also shown in vitro [67–69]. Surprisingly, anti-apoptotic effects of low concentrations of BPs were also reported in vitro and in vivo [70, 71]. In animal models, BPs generally reduced bone formation markers [21, 22, 27, 28, 33, 47, 72, 73] strongly indicating that either directly or indirectly they can affect bone cells other than osteoclasts.

Bone Marrow

Besides differential osteoblast and osteoclast induction potential, the composition of the bone marrow was also shown to differ among bone sites. Long bones contain more marrow fat than flat bones, and murine long-bone marrow contains more osteoclast precursors than jaw bone marrow [57]. Mesenchymal stem cells from the mandible have higher proliferation rates than those from long bones [74].

Despite its high specificity for bone-resorbing osteoclasts, BP uptake has also been noted in monocytes of rabbit bone marrow [75]. Monocyte/macrophage marker RNA expression in the bone marrow was also altered by ZA in rat long bones [76]. In vitro, non-bone cells cultured next to bone-resorbing osteoclasts were shown to internalize BP released from the bone [77]. In addition, the current authors showed that long-term treatment with ZA reduced the number of jaw bone marrow cells (unpublished results). Since macrophages are highly endocytic cells [78], they may be a major cell type in the bone marrow taking up BPs. Therefore, the effect of BPs on macrophage functioning would be an interesting direction for future research [79•, 80].

Collectively, the close interaction between different cell types in the bone microenvironment, and the effects that BPs can have on those cells, should all be considered when formulating a hypothesis explaining the pathophysiology of osteonecrosis of the jaw.

Hypotheses

Since both BPs and denosumab were shown to induce ONJ, the most comprehensive hypothesis would explain both relations and, at the same time, would also explain why ONJ is rare in other bones. Denosumab and BPs both inhibit osteoclasts and their activity, pointing towards suppression of bone turnover as a likely underlying mechanism. However, indirect effects through coupling with osteoblasts or other cells in the environment should not be ruled out. Coupling and direct effects on non-osteoclastic cells may be differently influenced by RANKL inhibition due to denosumab versus induction of apoptosis by BPs. Therefore, a direct comparison of denosumab and BPs on osteoclast numbers, osteoclast activity, osteoblast activity, and bone turnover in the jaws versus other bones would be of great interest. Although de Molon et al. [25••] did study the effect of both RANKL inhibitors and BPs in a mouse model of ONJ of the jaw, they did not assess the effects on other bones.

Suppression of Bone Turnover

Impaired osteoclast activity and the potential inability to heal microdamage may lead to an accumulation of necrotic bone tissue [40, 41]. This hypothesis does not explain though how a specific bone is exposed or why other bones are not affected. The effect may be stronger in the jaw if more BPs accumulate there, but that effect would not explain why denosumab is also related to ONJ. Also, in mice, we did not find any evidence that BPs more potently inhibit bone turnover in the jaw than in the long bones; on the contrary, it seemed to be the other way around (unpublished results).

Toxicity to Non-osteoclasts

A potentially toxic effect of BPs to other cell types in the bone may lead to necrosis and exposed bone. In vitro, BPs were indeed shown to inhibit proliferation of endothelial cells and angiogenesis [36, 81], reduce the wound healing capacity of murine epithelial cells [82], and furthermore, reduce the viability of periodontal ligament fibroblasts [83, 84] and macrophages [77, 85]. Osteoblasts and osteocytes were reported to be both positively and negatively affected by BPs, which was reviewed by Bellido and Plotkin [86], and they attributed the

different findings to the concentration of the drug. It is therefore uncertain whether the low concentration of BPs in vivo to which non-osteoclasts are exposed would cause the severe toxicity that could lead to ONJ. However, we previously showed that jaw bone marrow cells internalized more BPs than long-bone marrow cells in vitro [59•] and that ZA reduced the number of jaw bone marrow cells in vivo (unpublished results). Also, repeated administration of BPs was shown to replace bone-bound BPs, indicating that previously administered BPs can be released into the environment, increasing the local concentration [26]. BP release from the bone and a minor effect on the healing capacity might be enough to lead to the onset of ONJ in the event of an extraction or other trauma [11], which may be enhanced by an infection [35, 87]. Due to the constant pathogenic challenge in the jaw, this hypothesis would explain why especially the jaw is affected by osteonecrosis, yet it does not explain how denosumab could cause ONJ. Bone site-specific effects of RANKL inhibition may exist though, since the RANKL/OPG ratio was shown to be different in long-bone and jaw bone marrow [57].

Infection

Evidently, soft tissue toxicity and infection are two closely related phenomena possibly explaining the pathophysiology of ONJ. Unlike long bones, the jaw is constantly exposed to pathogens present in dental plaque, particularly the *Actinomyces* species commonly found in ONJ [88]. Furthermore, there is a high coexistence of ONJ and periodontitis [13], all of which implies a role for infection in ONJ. The ability to overcome infections may be inhibited by the negative effect of BPs on immune cells such as macrophages [79•]. In addition, the decreased pH associated with inflammation may enhance the release of BPs from adjacent bone, making them available for uptake by other cells [35], thereby inducing soft tissue destruction leading to necrotic bone exposure. Conversely, disruption of the mucosa to expose bone increases the risk of infection [15•, 34, 46]. Remarkably, Leclous et al. [87] found that a higher bone marrow inflammatory infiltrate correlated with more severe ONJ. Finally, since different osseous sites may have distinct compositions of immune cells, bone site-specific responses are likely to occur.

Similar to the ONJ-hypothesis of BP toxicity to other cells, the infection hypothesis explains the difference between bone sites, but does not explain how denosumab is related to ONJ. Yet, inhibition of RANKL may affect the immune cells expressing RANK, such as T cells, B cells, dendritic cells, and macrophages [89, 90]. The effect of denosumab on those immune cells in different bones requires further investigation.

Conclusion

The multifactorial pathogenesis of osteonecrosis of the jaw seems to rely on interactions between bone cells and their unique site-specific environment. These interactions and the superimposed effects of anti-resorptive drugs on specific bones require further investigation. Gaining additional insight into bone site-specific cells may help in the development of more specific, anti-resorptive drugs.

Compliance with Ethical Standards

Conflict of Interest Vincent Everts, Jenny Vermeer, and Greetje Renders declare that they have no conflict of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
 - Of major importance
1. Frith JC, Monkkenon J, Blackburn GM, Russell RG, Rogers MJ. Clodronate and liposome-encapsulated clodronate are metabolized to a toxic ATP analog, adenosine 5'-(beta, gamma-dichloromethylene) triphosphate, by mammalian cells in vitro. *J Bone Miner Res*. 1997;12(9):1358–67.
 2. Frith JC, Monkkenon J, Auriola S, Monkkenon H, Rogers MJ. The molecular mechanism of action of the antiresorptive and antiinflammatory drug clodronate: evidence for the formation in vivo of a metabolite that inhibits bone resorption and causes osteoclast and macrophage apoptosis. *Arthritis Rheum*. 2001;44(9):2201–10.
 3. Luckman SP, Hughes DE, Coxon FP, Graham R, Russell G, Rogers MJ. Nitrogen-containing bisphosphonates inhibit the mevalonate pathway and prevent post-translational prenylation of GTP-binding proteins, including Ras. *J Bone Miner Res*. 1998;13(4):581–9.
 4. van Beek E, Pieterman E, Cohen L, Lowik C, Papapoulos S. Farnesyl pyrophosphate synthase is the molecular target of nitrogen-containing bisphosphonates. *Biochem Biophys Res Commun*. 1999;264(1):108–11.
 5. Russell RG, Watts NB, Ebetino FH, Rogers MJ. Mechanisms of action of bisphosphonates: similarities and differences and their potential influence on clinical efficacy. *Osteoporos Int*. 2008;19(6):733–59.
 6. Marx RE. Pamidronate (Aredia) and zoledronate (Zometa) induced avascular necrosis of the jaws: a growing epidemic. *J Oral Maxillofac Surg*. 2003;61(9):1115–7.
 7. Migliorati CA, Schubert MM, Peterson DE, Seneda LM. Bisphosphonate-associated osteonecrosis of mandibular and maxillary bone: an emerging oral complication of supportive cancer therapy. *Cancer*. 2005;104(1):83–93.
 8. Ruggiero SL, Mehrotra B, Rosenberg TJ, Engroff SL. Osteonecrosis of the jaws associated with the use of bisphosphonates: a review of 63 cases. *J Oral Maxillofac Surg*. 2004;62(5):527–34.
 9. Abu-Id MH, Wamke PH, Gottschalk J, Springer I, Wiltfang J, Acil Y, et al. “Bis-phossoy jaws”—high and low risk factors for bisphosphonate-induced osteonecrosis of the jaw. *J Craniomaxillofac Surg*. 2008;36(2):95–103.
 10. Edwards BJ, Hellstein JW, Jacobsen PL, Kaltman S, Mariotti A, Migliorati CA. Updated recommendations for managing the care of patients receiving oral bisphosphonate therapy: an advisory statement from the American Dental Association Council on Scientific Affairs. *J Am Dent Assoc*. 2008;139(12):1674–7.
 11. Reid IR, Cornish J. Epidemiology and pathogenesis of osteonecrosis of the jaw. *Nat Rev Rheumatol*. 2012;8(2):90–6.
 12. Filleul O, Crompton E, Saussez S. Bisphosphonate-induced osteonecrosis of the jaw: a review of 2,400 patient cases. *J Cancer Res Clin Oncol*. 2010;136(8):1117–24.
 13. Marx RE, Sawatari Y, Fortin M, Broumand V. Bisphosphonate-induced exposed bone (osteonecrosis/osteopetrosis) of the jaws: risk factors, recognition, prevention, and treatment. *J Oral Maxillofac Surg*. 2005;63(11):1567–75.
 14. Khosla S, Burr D, Cauley J, Dempster DW, Ebeling PR, Felsenberg D, et al. Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. *J Bone Miner Res*. 2007;22(10):1479–91.
 15. Khan AA, Morrison A, Hanley DA, Felsenberg D, McCauley LK, O’Ryan F, et al. Diagnosis and management of osteonecrosis of the jaw: a systematic review and international consensus. *J Bone Miner Res*. 2015;30(1):3–23. **This paper describes a systematic review, defines the most recent international consensus about the definition of ONJ, and gives recommendations for diagnosis and management.**
 16. Ruggiero SL, Dodson TB, Assael LA, Landesberg R, Marx RE, Mehrotra B. American Association of Oral and Maxillofacial Surgeons position paper on bisphosphonate-related osteonecrosis of the jaw—2009 update. *Aust Endod J*. 2009;35(3):119–30.
 17. Ruggiero SL, Fantasia J, Carlson E. Bisphosphonate-related osteonecrosis of the jaw: background and guidelines for diagnosis, staging and management. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2006;102(4):433–41.
 18. Abtahi J, Agholme F, Aspenberg P. Prevention of osteonecrosis of the jaw by mucoperiosteal coverage in a rat model. *Int J Oral Maxillofac Surg*. 2013;42(5):632–6.
 19. Aghaloo TL, Kang B, Sung EC, Shoff M, Ronconi M, Gotcher JE, et al. Periodontal disease and bisphosphonates induce osteonecrosis of the jaws in the rat. *J Bone Miner Res*. 2011;26(8):1871–82.
 20. Aghaloo TL, Cheong S, Bezouglaia O, Kostenuik P, Atti E, Dry SM, et al. RANKL inhibitors induce osteonecrosis of the jaw in mice with periapical disease. *J Bone Miner Res*. 2014;29(4):843–54.
 21. Aguirre JI, Akhter MP, Kimmel DB, Pingel JE, Williams A, Jorgensen M, et al. Oncologic doses of zoledronic acid induce osteonecrosis of the jaw-like lesions in rice rats (*Oryzomys palustris*) with periodontitis. *J Bone Miner Res*. 2012;27(10):2130–43.
 22. Allen MR, Burr DB. Mandible matrix necrosis in beagle dogs after 3 years of daily oral bisphosphonate treatment. *J Oral Maxillofac Surg*. 2008;66(5):987–94.
 23. Allen MR, Chu TM, Ruggiero SL. Absence of exposed bone following dental extraction in beagle dogs treated with 9 months of high-dose zoledronic acid combined with dexamethasone. *J Oral Maxillofac Surg*. 2013;71(6):1017–26.
 24. Bi Y, Gao Y, Ehirchiou D, Cao C, Kikuiru T, Le A, et al. Bisphosphonates cause osteonecrosis of the jaw-like disease in mice. *Am J Pathol*. 2010;177(1):280–90.

25. de Molon RS, Cheong S, Bezouglaia O, Dry SM, Pirih F, Cirelli JA, et al. Spontaneous osteonecrosis of the jaws in the maxilla of mice on antiresorptive treatment: a novel ONJ mouse model. *Bone*. 2014;68:11–9. **This paper compares the effect of RANKL inhibitors and BPs in a mouse model of ONJ. It indicates that osteoclast inhibition and inflammation play a key role in ONJ. It also shows that RANKL inhibitors are more potent inhibitors of osteoclasts in the jaw than ZA.**
26. Hokugo A, Sun S, Park S, McKenna CE, Nishimura I. Equilibrium-dependent bisphosphonate interaction with crystalline bone mineral explains anti-resorptive pharmacokinetics and prevalence of osteonecrosis of the jaw in rats. *Bone*. 2013;53(1):59–68.
27. Howie RN, Borke JL, Kurago Z, Daoudi A, Cray J, Zakhary IE, et al. A model for osteonecrosis of the jaw with zoledronate treatment following repeated major trauma. *PLoS One*. 2015;10(7):e0132520.
28. Huja SS, Mason A, Fenell CE, Mo X, Hueni S, D'Atri AM, et al. Effects of short-term zoledronic acid treatment on bone remodeling and healing at surgical sites in the maxilla and mandible of aged dogs. *J Oral Maxillofac Surg*. 2011;69(2):418–27.
29. Kikuiiri T, Kim I, Yamaza T, Akiyama K, Zhang Q, Li Y, et al. Cell-based immunotherapy with mesenchymal stem cells cures bisphosphonate-related osteonecrosis of the jaw-like disease in mice. *J Bone Miner Res*. 2010;25(7):1668–79.
30. Kuroshima S, Go VA, Yamashita J. Increased numbers of nonattached osteoclasts after long-term zoledronic acid therapy in mice. *Endocrinology*. 2012;153(1):17–28.
31. Kuroshima S, Yamashita J. Chemotherapeutic and antiresorptive combination therapy suppressed lymphangiogenesis and induced osteonecrosis of the jaw-like lesions in mice. *Bone*. 2013;56(1):101–9.
32. Pautke C, Kreutzer K, Weitz J, Knodler M, Munzel D, Wexel G, et al. Bisphosphonate related osteonecrosis of the jaw: a minipig large animal model. *Bone*. 2012;51(3):592–9.
33. Takaoka K, Yamamura M, Nishioka T, Abe T, Tamaoka J, Segawa E, et al. Establishment of an animal model of bisphosphonate-related osteonecrosis of the jaws in spontaneously diabetic torii rats. *PLoS One*. 2015;10(12):e0144355.
34. Landesberg R, Woo V, Cremers S, Cozin M, Marolt D, Vunjak-Novakovic G, et al. Potential pathophysiological mechanisms in osteonecrosis of the jaw. *Ann N Y Acad Sci*. 2011;1218:62–79.
35. Otto S, Hafner S, Mast G, Tischler T, Volkmer E, Schieker M, et al. Bisphosphonate-related osteonecrosis of the jaw: is pH the missing part in the pathogenesis puzzle? *J Oral Maxillofac Surg*. 2010;68(5):1158–61.
36. Fournier P, Boissier S, Filleul S, Guglielmi J, Cabon F, Colombel M, et al. Bisphosphonates inhibit angiogenesis in vitro and testosterone-stimulated vascular regrowth in the ventral prostate in castrated rats. *Cancer Res*. 2002;62(22):6538–44.
37. Santini D, Vincenzi B, Dicuonzo G, Avvisati G, Massacesi C, Battistoni F, et al. Zoledronic acid induces significant and long-lasting modifications of circulating angiogenic factors in cancer patients. *Clin Cancer Res*. 2003;9(8):2893–7.
38. Arnett T. Regulation of bone cell function by acid-base balance. *Proc Nutr Soc*. 2003;62(2):511–20.
39. Sato M, Grasser W, Endo N, Akins R, Simmons H, Thompson DD, et al. Bisphosphonate action. Alendronate localization in rat bone and effects on osteoclast ultrastructure. *J Clin Invest*. 1991;88(6):2095–105.
40. Hoefert S, Schmitz I, Tannapfel A, Eufinger H. Importance of microcracks in etiology of bisphosphonate-related osteonecrosis of the jaw: a possible pathogenetic model of symptomatic and non-symptomatic osteonecrosis of the jaw based on scanning electron microscopy findings. *Clin Oral Investig*. 2010;14(3):271–84.
41. Mashiba T, Hirano T, Turner CH, Forwood MR, Johnston CC, Burr DB. Suppressed bone turnover by bisphosphonates increases microdamage accumulation and reduces some biomechanical properties in dog rib. *J Bone Miner Res*. 2000;15(4):613–20.
42. Nicoletti P, Carstos VM, Palaska PK, Shen Y, Floratos A, Zavras AI. Genomewide pharmacogenetics of bisphosphonate-induced osteonecrosis of the jaw: the role of RBMS3. *Oncologist*. 2012;17(2):279–87.
43. Sarasquete ME, Garcia-Sanz R, Marin L, Alcoceba M, Chillón MC, Balanzategui A, et al. Bisphosphonate-related osteonecrosis of the jaw is associated with polymorphisms of the cytochrome P450 CYP2C8 in multiple myeloma: a genome-wide single nucleotide polymorphism analysis. *Blood*. 2008;112(7):2709–12.
44. Aghaloo TL, Felsenfeld AL, Tetradis S. Osteonecrosis of the jaw in a patient on Denosumab. *J Oral Maxillofac Surg*. 2010;68(5):959–63.
45. Taylor KH, Middlefell LS, Mizen KD. Osteonecrosis of the jaws induced by anti-RANK ligand therapy. *Br J Oral Maxillofac Surg*. 2010;48(3):221–3.
46. Allam E, Allen M, Chu TM, Ghoneima A, Jack Windsor L. In vivo effects of zoledronic acid on oral mucosal epithelial cells. *Oral Dis*. 2011;17(3):291–7.
47. Allen MR, Kubek DJ, Burr DB. Cancer treatment dosing regimens of zoledronic acid result in near-complete suppression of mandible intracortical bone remodeling in beagle dogs. *J Bone Miner Res*. 2010;25(1):98–105.
48. Sasaki M, Matsuura T, Katafuchi M, Tokutomi K, Sato H. Higher contents of mineral and collagen but lower of hydroxylysine of collagen in mandibular bone compared with those of humeral and femoral bones in human. *J Hard Tissue Biol*. 2010;19(3):175–80.
49. van den Bos T, Speijer D, Bank RA, Bromme D, Everts V. Differences in matrix composition between calvaria and long bone in mice suggest differences in biomechanical properties and resorption: special emphasis on collagen. *Bone*. 2008;43(3):459–68.
50. Wen D, Qing L, Harrison G, Golub E, Akintoye SO. Anatomic site variability in rat skeletal uptake and desorption of fluorescently labeled bisphosphonate. *Oral Dis*. 2011;17(4):427–32.
51. Everts V, de Vries TJ, Helfrich MH. Osteoclast heterogeneity: lessons from osteopetrosis and inflammatory conditions. *Biochim Biophys Acta*. 2009;1792(8):757–65.
52. Henriksen K, Bollerslev J, Everts V, Karsdal MA. Osteoclast activity and subtypes as a function of physiology and pathology—implications for future treatments of osteoporosis. *Endocr Rev*. 2011;32(1):31–63.
53. Everts V, Korper W, Jansen DC, Steinfors J, Lammerse I, Heera S, et al. Functional heterogeneity of osteoclasts: matrix metalloproteinases participate in osteoclastic resorption of calvarial bone but not in resorption of long bone. *FASEB J*. 1999;13(10):1219–30.
54. Cackowski FC, Anderson JL, Patrene KD, Choksi RJ, Shapiro SD, Windle JJ, et al. Osteoclasts are important for bone angiogenesis. *Blood*. 2010;115(1):140–9.
55. Zenger S, Ek-Rylander B, Andersson G. Long bone osteoclasts display an augmented osteoclast phenotype compared to calvarial osteoclasts. *Biochem Biophys Res Commun*. 2010;394(3):743–9.
56. Jansen ID, Mardones P, Lecanda F, de Vries TJ, Recalde S, Hoeben KA, et al. Ae2(a, b)-deficient mice exhibit osteopetrosis of long bones but not of calvaria. *FASEB J*. 2009;23(10):3470–81.
57. de Souza Faroni AP, Schoenmaker T, Azari A, Katchburian E, Cerri PS, de Vries TJ, et al. Jaw and long bone marrows have a different osteoclastogenic potential. *Calcif Tissue Int*. 2011;88(1):63–74.
58. Chaichanasakul T, Kang B, Bezouglaia O, Aghaloo TL, Tetradis S. Diverse osteoclastogenesis of bone marrow from mandible versus long bone. *J Periodontol*. 2014;85(6):829–36.
59. Vermeer JA, Jansen ID, Marthi M, Coxon FP, McKenna CE, Sun S, et al. Jaw bone marrow-derived osteoclast precursors internalize more bisphosphonate than long-bone marrow precursors. *Bone*. 2013;57(1):242–51. **It is shown that jaw bone marrow cells internalize more bisphosphonates, but that after uptake, jaw cells are less sensitive to bisphosphonates than long-bone marrow cells.**

60. Aghaloo TL, Chaichanasakul T, Bezouglaia O, Kang B, Franco R, Dry SM, et al. Osteogenic potential of mandibular vs. long-bone marrow stromal cells. *J Dent Res*. 2010;89(11):1293–8.
61. Zhang P, Men J, Fu Y, Shan T, Ye J, Wu Y, et al. Contribution of SATB2 to the stronger osteogenic potential of bone marrow stromal cells from craniofacial bones. *Cell Tissue Res*. 2012;350(3):425–37.
62. Akintoye SO, Lam T, Shi S, Brahim J, Collins MT, Robey PG. Skeletal site-specific characterization of orofacial and iliac crest human bone marrow stromal cells in same individuals. *Bone*. 2006;38(6):758–68.
63. Matsubara T, Suardita K, Ishii M, Sugiyama M, Igarashi A, Oda R, et al. Alveolar bone marrow as a cell source for regenerative medicine: differences between alveolar and iliac bone marrow stromal cells. *J Bone Miner Res*. 2005;20(3):399–409.
64. Wan Q, Schoenmaker T, Jansen ID, Bian Z, de Vries TJ, Everts V. Osteoblasts of calvaria induce higher numbers of osteoclasts than osteoblasts from long bone. *Bone*. 2016;86:10–21.
65. Du Y, Jiang F, Liang Y, Wang Y, Zhou W, Pan Y, et al. The angiogenic variation of skeletal site-specific human BMSCs from same alveolar cleft patients: a comparative study. *J Mol Histol*. 2016;47(2):153–68.
66. Wang Y, Wan C, Deng L, Liu X, Cao X, Gilbert SR, et al. The hypoxia-inducible factor alpha pathway couples angiogenesis to osteogenesis during skeletal development. *J Clin Invest*. 2007;117(6):1616–26.
67. Cornish J, Bava U, Callon KE, Bai J, Naot D, Reid IR. Bone-bound bisphosphonate inhibits growth of adjacent non-bone cells. *Bone*. 2011;49(4):710–6.
68. Idris AI, Rojas J, Greig IR, Van't Hof RJ, Ralston SH. Aminobisphosphonates cause osteoblast apoptosis and inhibit bone nodule formation in vitro. *Calcif Tissue Int*. 2008;82(3):191–201.
69. Pozzi S, Vallet S, Mukherjee S, Cirstea D, Vaghela N, Santo L, et al. High-dose zoledronic acid impacts bone remodeling with effects on osteoblastic lineage and bone mechanical properties. *Clin Cancer Res*. 2009;15(18):5829–39.
70. Plotkin LI, Weinstein RS, Parfitt AM, Roberson PK, Manolagas SC, Bellido T. Prevention of osteocyte and osteoblast apoptosis by bisphosphonates and calcitonin. *J Clin Invest*. 1999;104(10):1363–74.
71. Plotkin LI, Lezcano V, Thostenson J, Weinstein RS, Manolagas SC, Bellido T. Connexin 43 is required for the anti-apoptotic effect of bisphosphonates on osteocytes and osteoblasts in vivo. *J Bone Miner Res*. 2008;23(11):1712–21.
72. Hayami T, Pickarski M, Wesolowski GA, McLane J, Bone A, Destefano J, et al. The role of subchondral bone remodeling in osteoarthritis: reduction of cartilage degeneration and prevention of osteophyte formation by alendronate in the rat anterior cruciate ligament transection model. *Arthritis Rheum*. 2004;50(4):1193–206.
73. Hornby SB, Evans GP, Hornby SL, Pataki A, Glatt M, Green JR. Long-term zoledronic acid treatment increases bone structure and mechanical strength of long bones of ovariectomized adult rats. *Calcif Tissue Int*. 2003;72(4):519–27.
74. Yamaza T, Ren G, Akiyama K, Chen C, Shi Y, Shi S. Mouse mandible contains distinctive mesenchymal stem cells. *J Dent Res*. 2011;90(3):317–24.
75. Roelofs AJ, Coxon FP, Ebetino FH, Lundy MW, Henneman ZJ, Nancollas GH, et al. Fluorescent risedronate analogues reveal bisphosphonate uptake by bone marrow monocytes and localization around osteocytes in vivo. *J Bone Miner Res*. 2010;25(3):606–16.
76. Yamashita J, Koi K, Yang DY, McCauley LK. Effect of zoledronate on oral wound healing in rats. *Clin Cancer Res*. 2011;17(6):1405–14.
77. Coxon FP, Thompson K, Roelofs AJ, Ebetino FH, Rogers MJ. Visualizing mineral binding and uptake of bisphosphonate by osteoclasts and non-resorbing cells. *Bone*. 2008;42(5):848–60.
78. Ryter A. Relationship between ultrastructure and specific functions of macrophages. *Comp Immunol Microbiol Infect Dis*. 1985;8(2):119–33.
79. Katsarelis H, Shah NP, Dhariwal DK, Pazianas M. Infection and medication-related osteonecrosis of the jaw. *J Dent Res*. 2015;94(4):534–9. **This review highlights the possible importance of macrophages and the immune system in the pathogenesis of ONJ.**
80. Pazianas M. Osteonecrosis of the jaw and the role of macrophages. *J Natl Cancer Inst*. 2011;103(3):232–40.
81. Wood J, Bonjean K, Ruetz S, Bellahcene A, Devy L, Foidart JM, et al. Novel antiangiogenic effects of the bisphosphonate compound zoledronic acid. *J Pharmacol Exp Ther*. 2002;302(3):1055–61.
82. Landesberg R, Cozin M, Cremers S, Woo V, Kousteni S, Sinha S, et al. Inhibition of oral mucosal cell wound healing by bisphosphonates. *J Oral Maxillofac Surg*. 2008;66(5):839–47.
83. Agis H, Blei J, Watzek G, Gruber R. Is zoledronate toxic to human periodontal fibroblasts? *J Dent Res*. 2010;89(1):40–5.
84. Correia Vde F, Caldeira CL, Marques MM. Cytotoxicity evaluation of sodium alendronate on cultured human periodontal ligament fibroblasts. *Dent Traumatol*. 2006;22(6):312–7.
85. Moreau MF, Guillet C, Massin P, Chevalier S, Gascan H, Basle MF, et al. Comparative effects of five bisphosphonates on apoptosis of macrophage cells in vitro. *Biochem Pharmacol*. 2007;73(5):718–23.
86. Bellido T, Plotkin LI. Novel actions of bisphosphonates in bone: preservation of osteoblast and osteocyte viability. *Bone*. 2011;49(1):50–5.
87. Lesclous P, Abi Najm S, Carrel JP, Baroukh B, Lombardi T, Willi JP, et al. Bisphosphonate-associated osteonecrosis of the jaw: a key role of inflammation? *Bone*. 2009;45(5):843–52.
88. Hansen T, Kunkel M, Springer E, Walter C, Weber A, Siegel E, et al. Actinomycosis of the jaws—histopathological study of 45 patients shows significant involvement in bisphosphonate-associated osteonecrosis and infected osteoradionecrosis. *Virchows Arch*. 2007;451(6):1009–17.
89. Ferrari-Lacraz S, Ferrari S. Do RANKL inhibitors (denosumab) affect inflammation and immunity? *Osteoporos Int*. 2011;22(2):435–46.
90. Toulis KA, Anastasilakis AD. Increased risk of serious infections in women with osteopenia or osteoporosis treated with denosumab. *Osteoporos Int*. 2010;21(11):1963–4.