



# Update on the Pathophysiology of Interstitial Cystitis /Bladder Pain Syndrome

Yoshiyuki Akiyama<sup>1</sup>

Published online: 6 January 2020  
© Springer Science+Business Media, LLC, part of Springer Nature 2020

## Abstract

**Purpose of Review** Interstitial cystitis/bladder pain syndrome (IC/BPS) is a chronic, potentially debilitating condition characterized by lower urinary tract symptoms and pain perceived to be related to the bladder. The etiology of IC/BPS has been rigorously studied for more than a century, but remains unknown. IC/BPS comprises a wide variety of clinical phenotypes with different potential etiologies. Recently, the importance of IC/BPS subtyping has become recognized. In this review, we revisit current hypotheses on IC/BPS pathophysiology and discuss the most likely causes of IC/BPS according to current research.

**Recent Findings** Recent histological and genomic analyses revealed that IC/BPS with Hunner lesions is a distinct inflammatory disorder characterized by epithelial denudation and frequent clonal expansion of infiltrating B cells, in association with biological processes involved in immune responses and infectious disease. Meanwhile, IC/BPS without Hunner lesions is an unrelated, non-inflammatory disorder with few histological changes, and which is potentially associated with systemic neurophysiological/endocrine abnormalities. Recent evidence has also cast doubt on the importance of features that have been conventionally considered significant in IC/BPS pathophysiology, such as mast cell infiltration or glomerulation.

**Summary** IC/BPS with Hunner lesions should be considered IC, and IC/BPS without Hunner lesions should be considered BPS. Clear and proper phenotyping of IC/BPS is necessary for the successful diagnosis and treatment of IC/BPS and to facilitate future research on IC/BPS pathophysiology.

**Keywords** Interstitial cystitis · Bladder pain syndrome · Pathophysiology · Infection · Immune response

## Introduction

Interstitial cystitis/bladder pain syndrome (IC/BPS) is a complex of chronic debilitating symptoms characterized by urologic chronic pelvic pain associated with lower urinary tract symptoms [1]. IC/BPS comprises a diverse variety of clinical phenotypes with different potential etiologies. However, precise subtyping of IC/BPS remains to be established due to its unknown etiology and the diverse sites and degrees of symptoms. Currently, one clear phenotype that has emerged is IC/BPS with Hunner lesions [the International Society for the Study of Bladder Pain Syndrome (ESSIC) criteria BPS type

3]. Recent evidence suggests that IC/BPS with Hunner lesions shows distinct features with respect to histology, gene expression, and prognosis compared with other forms of IC/BPS [2, 3•, 4]. IC/BPS with Hunner lesions is a robust inflammatory disorder characterized by epithelial denudation, pancystitis, and frequent clonal expansion of infiltrating B cells, with overexpression of genes involved in immune responses and infection [3•, 5•]. Meanwhile, IC/BPS without Hunner lesions is a distinct non-inflammatory disorder characterized by preservation of the urothelium layer and symptom spread beyond the bladder [2, 6•]. Thus, IC/BPS with and without Hunner lesions should be clearly distinguished, and the two should not be combined when evaluating the potential underlying pathophysiology or when designing clinical trials. Experts in the US emphasized this notion recently and suggested that past clinical trial failures and the persistent inconclusive results of basic research on IC/BPS might be attributable to the lack of proper phenotyping [7]. In this review, the potential etiologies of IC/BPS will be discussed with respect to IC/BPS phenotype.

This article is part of the *Topical Collection on BPS/Interstitial Cystitis*

✉ Yoshiyuki Akiyama  
yakiyamauro-ky@umin.ac.jp

<sup>1</sup> Department of Urology, Graduate School of Medicine, The University of Tokyo, 3-1, 7 Chome, Hongo, Bunkyo-ku, Tokyo, Japan

## Potential Etiologies of IC/BPS

### Urothelium Defects

Urothelial deficiency is generally considered a primary cause of IC/BPS [8]. Functional and/or quantitative loss of urothelial barrier function permits urinary subnoxious irritants to directly contact the suburothelial layer, resulting in inflammatory responses or afferent nerve stimulation [9].

As mentioned above, it is likely that quantitative loss of the urothelium plays a role in the etiology of IC/BPS with Hunner lesions, in which the urothelium is almost completely denuded at the lesion sites [2]. However, epithelial functional loss may occur in IC/BPS with and without Hunner lesions. The detailed mechanisms thought to contribute to this urothelium deficiency are as follows.

#### 1. Abnormality of the glycosaminoglycan (GAG) layer of the urothelium

Parsons and colleagues suggested that abnormalities of the GAG layer might be associated with the urothelial dysfunction of IC/BPS [8]. The GAG layer, which consists of glycoproteins and proteoglycans, plays a pivotal role in protective barrier function [10]. Disruption of the GAG layer increases the permeability of the urothelial layer, leading to subepithelial inflammation and lower urinary tract hypersensitivity symptoms such as bladder/pelvic pain and urinary frequency [10]. The use of conventional intravesical therapies with GAG family components such as heparan sulfate, chondroitin sulfate, or hyaluronic acid is based on the rationale that replenishment of the GAG layer might promote urothelial layer recovery [11].

#### 2. Impaired cell adhesion

Liu et al. reported that tight junction proteins such as E-cadherin and zonula occludens-1 (ZO-1) are down-regulated in patients with IC/BPS compared with control patients or those with overactive bladder syndrome [12]. Interestingly, the subjects analyzed in that study all had IC/BPS without Hunner lesions. Montalbetti et al. demonstrated that, in rats, increased urothelial permeability caused by disrupted urothelial tight junctions led to bladder afferent nerve hyperexcitability and upstream central nerve sensitization with slight inflammatory changes, without altering epithelial quantity or umbrella cell polarity/differentiation [9, 13]. Impaired cell adhesion and increased urothelial permeability (e.g., not quantitative but functional disruption of the urothelium) may be associated with the pathophysiology of IC/BPS, especially IC/BPS without Hunner lesions.

#### 3. Aberrant epithelial proliferation

Loss of epithelial cells due to altered production of antiproliferative factors or certain growth factors or increased apoptotic activity has been reported in IC/BPS [14, 15]. This hypothesis would implicate quantitative loss of epithelium in the etiology of IC/BPS with Hunner lesions.

#### 4. Toxic substances in the urine

Parsons et al. suggested that cationic urinary components could potentially be cytotoxic to urothelial cells, leading to bladder mucosal injury [16]. It is also well known that ketamine-induced cystitis resembles IC/BPS with Hunner lesions both histologically and clinically [17]. Although the detailed etiology of ketamine cystitis has not been clearly elucidated, direct damage to the urothelium by urine metabolites of ketamine has been suggested as a possible mechanism [17]. In IC/BPS with Hunner lesions, urine itself also seems to exacerbate the hypersensitivity symptoms by leaking into the bladder subepithelial layer at the lesion sites, where the full layer of urothelium is frequently sloughed off [2]. It is well recognized that there is an association between diet and symptom severity in IC/BPS [18–20]. In routine clinical practice, we find that patients with IC/BPS with Hunner lesions often notice a relationship between intake of specific foods and worsening symptoms. Alterations in the osmolarity, composition, or acidity of the urine due to the consumption of specific foods may stimulate the peripheral afferent nerves in the bladder, in addition to the direct cytotoxic effects of dietary metabolites, medications, or dietary supplements on the urothelium [21–23]. Given this aspect of symptom triggering in IC/BPS, the responsiveness of the Hunner lesion-targeted therapies is reasonable. [4]

#### 5. Autoimmunity against the bladder urothelium

Specific autoantibodies have not been identified in IC/BPS, although non-specific anti-nuclear antibodies have been reported in some cases [24–27]. However, the clinical manifestations of patients with bladder disorders associated with other systemic autoimmune diseases such as Sjogren's syndrome, systemic lupus erythematosus, or autoimmune thyroiditis are quite similar to those of patients with IC/BPS with Hunner lesions [28, 29]. Deposits of immunoglobulin and complement are observed in the bladders of patients with systemic autoimmune disorders with urinary symptoms as well as in patients with IC/BPS [27, 30, 31]. In patients with IC/BPS with Hunner lesions, epithelial denudation and lymphoplasmacytic infiltration of the bladder are observed, with frequent clonal expansion of infiltrating B cells [5]. These findings strongly imply that autoimmune responses underlie the pathophysiology of IC/BPS with Hunner lesions. In

an animal model of autoimmune cystitis induced by immunization with uroplakin and in transgenic mouse model that expresses antigenic ovalbumin (OVA) in the urothelium and OVA-specific receptor on CD8-positive T cells, mice spontaneously develop a form of cystitis that strongly resembles IC/BPS with Hunner lesions in histology and lower urinary tract hypersensitivity symptoms [32–34].

## Inflammation

### Activation of Mast Cells

Mast cell infiltration has been also considered a characteristic feature of IC/BPS [35–41]. Before the year 2000, Giemsa staining, toluidine blue staining, periodic acid-Schiff staining, and immunohistochemistry against c-kit were routinely used to identify mast cells. However, these staining methods are not specific for human mast cells [36, 40, 41]. In the 2000s, when an anti-tryptase antibody specific for human mast cells was developed [42], growing evidence cast doubt on the significance of mast cell infiltration in IC/BPS [43, 44••, 45••]. For example, we compared mast cell densities in the bladders of IC/BPS patients (with and without Hunner lesions) with those of non-IC controls (non-IC cystitis and normal bladder, respectively). A similar degree of background lymphoplasmacytic infiltration was observed in each pair (IC/BPS with Hunner lesions vs. non-IC cystitis, or IC/BPS without Hunner lesions vs. normal bladder). These results indicated that mast cell densities did not differ between IC/BPS and controls with equal background inflammation [45••], which suggests that mast cell infiltration may not be a specific histological feature of IC/BPS. In addition to the quantitative analysis, functional assessment of mast cells in IC/BPS should be performed in order to more firmly establish the role of mast cell infiltration in IC/BPS.

### Immunologic Inflammation

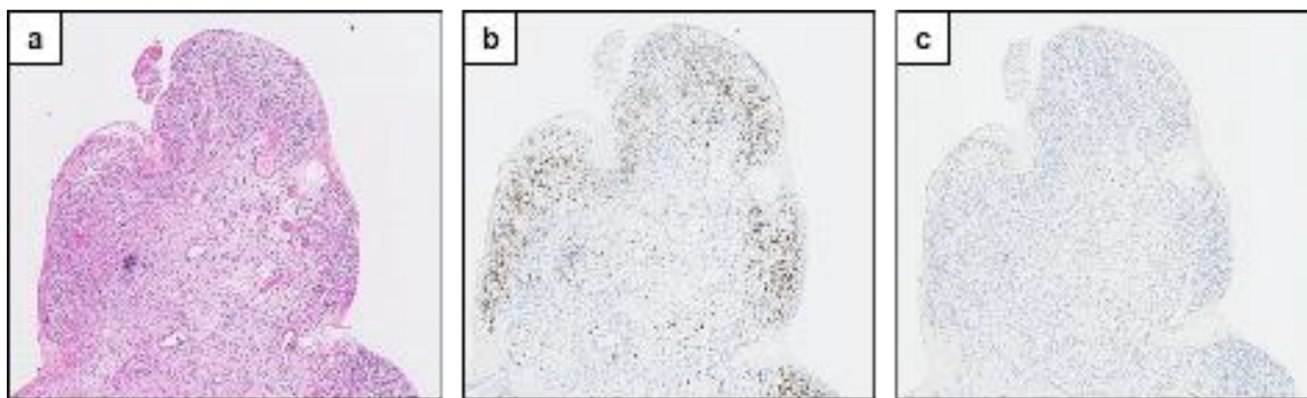
Deposits of immunoglobulin and complement, and increased expression of the chemokine receptor CXCR3 and its ligands (CXCL9, 10, and 11), have been reported in the bladders of IC/BPS with Hunner lesions [27, 31, 46, 47]. Recently, we found that frequent expansion of light chain-restricted B cells was observed in the bladders of IC/BPS patients with Hunner lesions (Fig. 1) [5••]. This evidence collectively suggests that specific immune responses such as autoimmune responses may underlie the pathophysiology of IC/BPS with Hunner lesions.

In this context, the higher prevalence of comorbid autoimmune disorders in patients with IC/BPS with Hunner lesions may reflect this disease property [48, 49]. Potential pathogenic antigens, e.g., self-antigens in the bladder tissue or exogenous antigens such as those derived from microorganisms or chemical substances in the urine, are worth evaluating in the future.

Our recent whole transcriptome analysis indicated that biological pathways related to the immune system and infectious disease were significantly enriched in IC/BPS with Hunner lesions [3••]. These results suggest that autoimmunity in conjunction with infection may underlie the pathophysiology of IC/BPS with Hunner lesions. However, specific autoantibodies have not been identified in IC/BPS, as mentioned above. Furthermore, the inefficacy of antibiotics and the lack of detection of pathogenic microorganisms by urine culture or bacterial DNA sequencing have cast doubt on bacterial infection as a potential etiologic factor in IC/BPS [50–53]. On the other hand, it has been reported that female IC patients had a higher prevalence of positive urine cultures [50, 54] and alterations in the diversity of urine bacterial flora [55, 56]. Patients with IC/BPS with Hunner lesions also had a higher frequency of Epstein-Barr virus infection [57], suggesting that microorganism infection might play a role in the pathogenesis of IC/BPS. Further studies on the relationship between immunologic inflammation frequently accompanied by B cell clonal expansion and microorganism infection may yield clues into the pathophysiology of IC/BPS.

### Neurogenic Inflammation

Neurochemical transmitters can stimulate the afferent nerves or induce stromal inflammatory changes such as fibrosis or edema in the bladder. This process is mainly mediated by mast cells. Granules released by mast cells containing inflammatory mediators such as histamine, serotonin, tryptase, tumor necrosis factor (TNF)- $\alpha$ , or nerve growth factor (NGF) can stimulate the peripheral nerves in the bladder mucosa, which then release neuropeptides such as substance P and calcitonin gene-related peptide (CGRP) that further exacerbate mast cell degranulation (the nerve-mast cell interaction axis) [58–66]. As a consequence, chronic stimulation of the afferent nerves could lead to altered neural plasticity or central nerve sensitization in the dorsal root ganglia or the upper spinal cord, which may contribute to symptom persistence in IC/BPS [67]. Chitinase-like protein (YKL-40), another protein contained within mast cell granules, promotes stromal edema and fibrosis [68]. Mast cells are involved in diverse pathophysiological processes, such as synthesis of proinflammatory cytokines, recruitment of leucocytes, and vascular remodeling [69]. However, again, recent evidence has cast doubt on the significance of mast cells in IC/BPS pathophysiology. Gamper et al. reported that the degree of mast cell degranulation as well as mast cell numbers did not differ significantly between patients with IC/BPS and overactive bladder and non-IC controls without bladder pain [44••]. The role of neurogenic inflammation in IC/BPS should be revisited to take the proper IC/BPS phenotyping and control setting into account.



**Fig. 1** Light chain restriction in infiltrating plasma cells in the bladder of IC/BPS with Hunner lesions. **a** Hematoxylin and eosin staining shows dense inflammatory infiltrates in the subepithelial layer. **b, c** In situ

hybridization for the kappa chain (**b**) or lambda chain (**c**). Most plasma cells express the kappa chain

### Increased Nociceptive Reflux Pathways

Increased gene expression of transient receptor potential channels (TRPs) and certain chemokines has been reported in IC/BPS with Hunner lesions [70]. These findings may reflect the increased activation of nociceptive reflux pathways in IC/BPS, possibly as a result of chronic inflammatory processes. On the other hand, the severity of symptoms in IC/BPS without Hunner lesions is similar to that in IC/BPS with Hunner lesions despite the absence of bladder inflammation. Other aberrant sensory pathways that could lead to central sensitization may underlie the pathophysiology of IC/BPS without Hunner lesions.

### Increased Angiogenesis in the Urothelium

Increased angiogenesis is considered another pathogenic feature of IC/BPS [71]. Glomerulation (mucosal bleeding after bladder overdistension), which is considered a specific diagnostic marker of IC/BPS, could be associated with increased angiogenesis in conjunction with epithelial dysfunction [72, 73]. The angiogenetic factor most frequently reported to be increased in IC/BPS is vascular endothelial growth factor (VEGF), a proinflammatory growth factor that induces neo-vascularization and is involved in the pathogenesis of many chronic inflammatory disorders [74–76]. The elevated VEGF expression in IC/BPS with Hunner lesions is therefore likely a consequence of chronic inflammatory responses. However, our recent whole transcriptome study and quantitative microvasculature analysis failed to detect any significant difference in gene expression profile, biological pathways, microvessel density, VEGF expression, or subepithelial inflammation in patients with IC/BPS without Hunner lesions, regardless of the presence or absence of glomerulations [3••]. Further, a recent study questioned the connection between glomerulations and IC/BPS [77•]. Although it cannot be

completely ruled out, the role of glomerulations in IC/BPS pathophysiology is currently unclear.

### IC/BPS Without Hunner Lesions as a Potential Somatoform Disorder

A recent study reported that increased bladder capacity was associated with a set of psychological factors such as dissociative pathology and childhood relational trauma in patients with IC/BPS. These psychological factors are thought to underlie somatoform disorders and functional somatic syndromes (FSSs) [78, 79]. It is well known that patients with IC/BPS without Hunner lesions usually retain their bladder capacity significantly larger than those with Hunner lesions, and their symptoms frequently extend systemically (beyond the bladder) [6•, 80••]. Chen et al. indicated that somatic symptoms could potentially be linked to biological pathways that increase the risk of IC/BPS without Hunner lesions [81]. Warren et al. pointed out that there is strong overlap between the symptoms and comorbidities of IC/BPS without Hunner lesions and those of FSSs, including irritable bowel syndrome, fibromyalgia, chronic fatigue syndrome, and migraines [80••]. Patients with IC/BPS without Hunner lesions have all the characteristics and meet all the proposed criteria of a FSS [82••]. This evidence suggests that IC/BPS without Hunner lesions may share its neurophysiological process with widely recognized FSSs such as chronic fatigue syndrome, irritable bowel syndrome, and fibromyalgia, which lead to central sensitization [83]. The underlying pathophysiology of FSSs remains unclear, but aberrant neuroimmune or endocrine processes with certain stressors have been suggested [84]. A multidisciplinary approach (behavioral, biomedical, and cognitive) is appropriate for the treatment of IC/BPS without Hunner lesions.



## Conclusions

Recent growing evidence has changed the current thinking on IC/BPS. IC/BPS with Hunner lesions is an inflammatory disease potentially associated with immune responses and infection, and should be considered distinct from the broad IC/BPS syndrome umbrella. Meanwhile, IC/BPS without Hunner lesions is a non-inflammatory disorder that is often associated with systemic hypersensitivity, potentially due to neurophysiological/endocrine abnormalities. Few bladder histological findings are noted in IC/BPS without Hunner lesions. This subtype may share the pathogenesis of widely recognized FSSs. Clear and proper phenotyping of IC/BPS will improve its clinical management, as the successful treatment of IC/BPS with Hunner lesions (IC) could be dependent on Hunner lesion-targeted therapies, while the treatment of IC/BPS without Hunner lesions (BPS) could require strategies similar to those used for the treatment of other somatic syndromes. Considering IC and BPS patients separately will also facilitate future research on IC/BPS.

## Compliance with Ethical Standards

**Conflict of Interest** The author declares that there are no conflicts of interest.

**Human and Animal Rights and Informed Consent** All reported studies/experiments with human or animal subjects performed by the authors have been previously published and complied with all applicable ethical standards (including the Helsinki declaration and its amendments, institutional/national research committee standards, and international/national/institutional guidelines).

## References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Hanno PM, Erickson D, Moldwin R, Faraday MM, American Urological A. Diagnosis and treatment of interstitial cystitis/bladder pain syndrome: AUA guideline amendment. *J Urol*. 2015;193(5):1545–53. <https://doi.org/10.1016/j.juro.2015.01.086>.
2. Akiyama Y, Homma Y, Maeda D. Pathology and terminology of interstitial cystitis/bladder pain syndrome: a review. *Histol Histopathol*. 2019;34:25–32. <https://doi.org/10.14670/HH-18-028>.
3. Akiyama Y, Maeda D, Katoh H, Morikawa T, Niimi A, Nomiya A, et al. Molecular taxonomy of interstitial cystitis/bladder pain syndrome based on whole transcriptome profiling by next-generation RNA sequencing of bladder mucosal biopsies. *J Urol*. 2019;202:290–300. <https://doi.org/10.1097/JU.000000000000234> **Whole transcriptome and quantitative microvasculature analyses revealed distinct gene expression and microvasculature in IC/BPS with Hunner lesions.**
4. Chennamsetty A, Khouradji I, Goike J, Killinger KA, Girdler B, Peters KM. Electrosurgical management of Hunner ulcers in a referral center's interstitial cystitis population. *Urology*. 2015;85(1):74–8. <https://doi.org/10.1016/j.urology.2014.09.012>.
5. Maeda D, Akiyama Y, Morikawa T, Kunita A, Ota Y, Katoh H, et al. Hunner-type (classic) interstitial cystitis: a distinct inflammatory disorder characterized by pancystitis, with frequent expansion of clonal B-cells and epithelial denudation. *PLoS One*. 2015;10(11):e0143316. <https://doi.org/10.1371/journal.pone.0143316> **Quantitative histological analysis revealed epithelial denudation and B cell clonal expansion as distinct and crucial histological features of IC/BPS with Hunner lesions.**
6. Peters KM, Killinger KA, Mounayer MH, Boura JA. Are ulcerative and nonulcerative interstitial cystitis/painful bladder syndrome 2 distinct diseases? A study of coexisting conditions. *Urology*. 2011;78(2):301–8. <https://doi.org/10.1016/j.urology.2011.04.030> **Distinct clinical characteristics between IC/BPS subtypes were demonstrated.**
7. Nickel JC, Moldwin R, Hanno P, Dmochowski R, Peters KM, Payne C, et al. Targeting the SHIP1 pathway fails to show treatment benefit in interstitial cystitis/bladder pain syndrome: lessons learned from evaluating potentially effective therapies in this enigmatic syndrome. *J Urol*. 2019;202(2):301–8. <https://doi.org/10.1097/JU.000000000000192>.
8. Parsons CL, Lilly JD, Stein P. Epithelial dysfunction in nonbacterial cystitis (interstitial cystitis). *J Urol*. 1991;145(4):732–5. [https://doi.org/10.1016/s0022-5347\(17\)38437-9](https://doi.org/10.1016/s0022-5347(17)38437-9).
9. Montalbetti N, Rued AC, Taiclet SN, Birder LA, Kullmann FA, Carattino MD. Urothelial tight junction barrier dysfunction sensitizes bladder afferents. *eNeuro*. 2017;4(3):ENEURO.0381-16. <https://doi.org/10.1523/ENEURO.0381-16.2017>.
10. Klingler CH. Glycosaminoglycans: how much do we know about their role in the bladder? *Urologia*. 2016;83(Suppl 1):11–4. <https://doi.org/10.5301/uro.5000184>.
11. Janssen DA, van Wijk XM, Jansen KC, van Kuppevelt TH, Heesakkers JP, Schalken JA. The distribution and function of chondroitin sulfate and other sulfated glycosaminoglycans in the human bladder and their contribution to the protective bladder barrier. *J Urol*. 2013;189(1):336–42. <https://doi.org/10.1016/j.juro.2012.09.022>.
12. Liu HT, Shie JH, Chen SH, Wang YS, Kuo HC. Differences in mast cell infiltration, E-cadherin, and zonula occludens-1 expression between patients with overactive bladder and interstitial cystitis/bladder pain syndrome. *Urology*. 2012;80(1):225 e13–8. <https://doi.org/10.1016/j.urology.2012.01.047>.
13. Montalbetti N, Rued AC, Clayton DR, Ruiz WG, Bastacky SI, Prakasam HS, et al. Increased urothelial paracellular transport promotes cystitis. *Am J Physiol Ren Physiol*. 2015;309(12):F1070–81. <https://doi.org/10.1152/ajprenal.00200.2015>.
14. Keay S, Kleinberg M, Zhang CO, Hise MK, Warren JW. Bladder epithelial cells from patients with interstitial cystitis produce an inhibitor of heparin-binding epidermal growth factor-like growth factor production. *J Urol*. 2000;164(6):2112–8.
15. Keay S, Seillier-Moisewitsch F, Zhang CO, Chai TC, Zhang J. Changes in human bladder epithelial cell gene expression associated with interstitial cystitis or antiproliferative factor treatment. *Physiol Genomics*. 2003;14(2):107–15. <https://doi.org/10.1152/physiolgenomics.00055.2003>.
16. Parsons CL, Bautista SL, Stein PC, Zupkas P. Cyto-injury factors in urine: a possible mechanism for the development of interstitial cystitis. *J Urol*. 2000;164(4):1381–4.
17. Jhang JF, Hsu YH, Kuo HC. Possible pathophysiology of ketamine-related cystitis and associated treatment strategies. *Int J*

- Urol : official journal of the Japanese Urological Association. 2015;22(9):816–25. <https://doi.org/10.1111/iju.12841>.
18. Gillespie L. Metabolic appraisal of the effects of dietary modification on hypersensitive bladder symptoms. *Br J Urol*. 1993;72(3):293–7.
  19. Friedlander JI, Shorter B, Moldwin RM. Diet and its role in interstitial cystitis/bladder pain syndrome (IC/BPS) and comorbid conditions. *BJU Int*. 2012;109(11):1584–91. <https://doi.org/10.1111/j.1464-410X.2011.10860.x>.
  20. Bassaly R, Downes K, Hart S. Dietary consumption triggers in interstitial cystitis/bladder pain syndrome patients. *Female Pelvic Med Reconstr Surg*. 2011;17(1):36–9. <https://doi.org/10.1097/SPV.0b013e3182044b5c>.
  21. Whitmore KE. Self-care regimens for patients with interstitial cystitis. *Urol Clin N Am*. 1994;21(1):121–30.
  22. Koziol JA, Clark DC, Gittes RF, Tan EM. The natural history of interstitial cystitis: a survey of 374 patients. *J Urol*. 1993;149(3):465–9. [https://doi.org/10.1016/s0022-5347\(17\)36120-7](https://doi.org/10.1016/s0022-5347(17)36120-7).
  23. Parsons CL, Shaw T, Berecz Z, Su Y, Zupkas P, Argade S. Role of urinary cations in the aetiology of bladder symptoms and interstitial cystitis. *BJU Int*. 2014;114(2):286–93. <https://doi.org/10.1111/bju.12603> **This report suggested urine as a potential causative factor of urothelial damage in IC/BPS.**
  24. Oravisto KJ. Interstitial cystitis as an autoimmune disease A review. *Eur Urol*. 1980;6(1):10–3.
  25. Silk MR. Bladder antibodies in interstitial cystitis. *J Urol*. 1970;103(3):307–9.
  26. Jokinen EJ, Alfthan OS, Oravisto KJ. Antitissue antibodies in interstitial cystitis. *Clin Exp Immunol*. 1972;11(3):333–9.
  27. Anderson JB, Parivar F, Lee G, Wallington TB, MacIver AG, Bradbrook RA, et al. The enigma of interstitial cystitis—an autoimmune disease? *Br J Urol*. 1989;63(1):58–63.
  28. Haarala M, Alanen A, Hietarinta M, Kiilholma P. Lower urinary tract symptoms in patients with Sjogren's syndrome and systemic lupus erythematosus. *Int Urogynecol J Pelvic Floor Dysfunct*. 2000;11(2):84–6.
  29. Leppilahti M, Tammela TL, Huhtala H, Kiilholma P, Leppilahti K, Auvinen A. Interstitial cystitis-like urinary symptoms among patients with Sjogren's syndrome: a population-based study in Finland. *Am J Med*. 2003;115(1):62–5.
  30. Boye E, Morse M, Huttner I, Erlanger BF, MacKinnon KJ, Klassen J. Immune complex-mediated interstitial cystitis as a major manifestation of systemic lupus erythematosus. *Clin Immunol Immunopathol*. 1979;13(1):67–76.
  31. Mattila J, Linder E. Immunoglobulin deposits in bladder epithelium and vessels in interstitial cystitis: possible relationship to circulating anti-intermediate filament autoantibodies. *Clin Immunol Immunopathol*. 1984;32(1):81–9.
  32. Izgi K, Altuntas CZ, Bicer F, Ozer A, Sakalar C, Li X, et al. Uroplakin peptide-specific autoimmunity initiates interstitial cystitis/painful bladder syndrome in mice. *PLoS One*. 2013;8(8):e72067. <https://doi.org/10.1371/journal.pone.0072067>.
  33. Altuntas CZ, Daneshgari F, Sakalar C, Goksoy E, Gulen MF, Kavran M, et al. Autoimmunity to uroplakin II causes cystitis in mice: a novel model of interstitial cystitis. *Eur Urol*. 2012;61(1):193–200. <https://doi.org/10.1016/j.eururo.2011.06.028>.
  34. Liu W, Evanoff DP, Chen X, Luo Y. Urinary bladder epithelium antigen induces CD8+ T cell tolerance, activation, and autoimmune response. *J Immunol*. 2007;178(1):539–46. <https://doi.org/10.4049/jimmunol.178.1.539>.
  35. Kastrop J, Hald T, Larsen S, Nielsen VG. Histamine content and mast cell count of detrusor muscle in patients with interstitial cystitis and other types of chronic cystitis. *Br J Urol*. 1983;55(5):495–500.
  36. Aldenborg F, Fall M, Enerback L. Proliferation and transepithelial migration of mucosal mast cells in interstitial cystitis. *Immunology*. 1986;58(3):411–6.
  37. Lynes WL, Flynn SD, Shortliffe LD, Lemmers M, Zipser R, Roberts LJ 2nd, et al. Mast cell involvement in interstitial cystitis. *J Urol*. 1987;138(4):746–52.
  38. Christmas TJ, Rode J. Characteristics of mast cells in normal bladder, bacterial cystitis and interstitial cystitis. *Br J Urol*. 1991;68(5):473–8.
  39. Theoharides TC, Sant GR, el Mansoury M, Letourneau R, Ucci AA Jr, Meares EM Jr. Activation of bladder mast cells in interstitial cystitis: a light and electron microscopic study. *J Urol*. 1995;153(3 Pt 1):629–36.
  40. Pecker R, Enerback L, Fall M, Aldenborg F. Recruitment, distribution and phenotypes of mast cells in interstitial cystitis. *J Urol*. 2000;163(3):1009–15.
  41. Yamada T, Murayama T, Mita H, Akiyama K. Subtypes of bladder mast cells in interstitial cystitis. *Int J Urol : official journal of the Japanese Urological Association*. 2000;7(8):292–7.
  42. Larsen MS, Mortensen S, Nordling J, Horn T. Quantifying mast cells in bladder pain syndrome by immunohistochemical analysis. *BJU Int*. 2008;102(2):204–7; discussion 7. <https://doi.org/10.1111/j.1464-410X.2008.07576.x>.
  43. Liu H-T, Jiang Y-H, Kuo H-C. Alteration of urothelial inflammation, apoptosis, and junction protein in patients with various bladder conditions and storage bladder symptoms suggest common pathway involved in underlying pathophysiology. *LUTS: Lower Urinary Tract Symptoms*. 2015;7(2):102–7. <https://doi.org/10.1111/luts.12062>.
  44. Gamper M, Regauer S, Welter J, Eberhard J, Viereck V. Are mast cells still good biomarkers for bladder pain syndrome/interstitial cystitis? *J Urol*. 2015;193(6):1994–2000. <https://doi.org/10.1016/j.juro.2015.01.036> **This study revealed the potential insignificance of mast cells in IC/BPS.**
  45. Akiyama Y, Maeda D, Morikawa T, Niimi A, Nomiya A, Yamada Y, et al. Digital quantitative analysis of mast cell infiltration in interstitial cystitis. *Neurourol Urodyn*. 2018;37(2):650–7. <https://doi.org/10.1002/nau.23365> **This study revealed the potential insignificance of mast cells in IC/BPS.**
  46. Ogawa T, Homma T, Igawa Y, Seki S, Ishizuka O, Imamura T, et al. CXCR3 binding chemokine and TNFSF14 over expression in bladder urothelium of patients with ulcerative interstitial cystitis. *J Urol*. 2010;183(3):1206–12. <https://doi.org/10.1016/j.juro.2009.11.007>.
  47. Akiyama Y, Morikawa T, Maeda D, Shintani Y, Niimi A, Nomiya A, et al. Increased CXCR3 expression of infiltrating plasma cells in Hunner type interstitial cystitis. *Sci Rep*. 2016;6:28652. <https://doi.org/10.1038/srep28652>.
  48. Pecker R, Atanasiu L, Logadottir Y. Intercurrent autoimmune conditions in classic and non-ulcer interstitial cystitis. *Scand J Urol Nephrol*. 2003;37(1):60–3. <https://doi.org/10.1080/0036590310008721>.
  49. van de Merwe JP. Interstitial cystitis and systemic autoimmune diseases. *Nat Clin Pract Urol*. 2007;4(9):484–91. <https://doi.org/10.1038/ncpuro0874>.
  50. Keay S, Schwalbe RS, Trifillis AL, Lovchik JC, Jacobs S, Warren JW. A prospective study of microorganisms in urine and bladder biopsies from interstitial cystitis patients and controls. *Urology*. 1995;45(2):223–9.
  51. Keay S, Zhang CO, Baldwin BR, Jacobs SC, Warren JW. Polymerase chain reaction amplification of bacterial 16S rRNA genes in interstitial cystitis and control patient bladder biopsies. *J Urol*. 1998;159(1):280–3. [https://doi.org/10.1016/s0022-5347\(01\)64082-5](https://doi.org/10.1016/s0022-5347(01)64082-5).

52. Al-Hadithi HN, Williams H, Hart CA, Frazer M, Adams EJ, Richmond DH, et al. Absence of bacterial and viral DNA in bladder biopsies from patients with interstitial cystitis/chronic pelvic pain syndrome. *J Urol*. 2005;174(1):151–4. <https://doi.org/10.1097/01.ju.0000161605.14804.a9>.
53. Hanno PM. Diagnosis of interstitial cystitis. *Urol Clin N Am*. 1994;21(1):63–6.
54. Haarala M, Kiilholma P, Lehtonen OP. Urinary bacterial flora of women with urethral syndrome and interstitial cystitis. *Gynecol Obstet Investig*. 1999;47(1):42–4. <https://doi.org/10.1159/000010060>.
55. Siddiqui H, Lagesen K, Nederbragt AJ, Jeansson SL, Jakobsen KS. Alterations of microbiota in urine from women with interstitial cystitis. *BMC Microbiol*. 2012;12:205. <https://doi.org/10.1186/1471-2180-12-205>.
56. Abernethy MG, Rosenfeld A, White JR, Mueller MG, Lewicky-Gaupp C, Kenton K. Urinary microbiome and cytokine levels in women with interstitial cystitis. *Obstet Gynecol*. 2017;129(3):500–6. <https://doi.org/10.1097/AOG.0000000000001892>.
57. Jhang JF, Hsu YH, Peng CW, Jiang YH, Ho HC, Kuo HC. Epstein-Barr virus as a potential etiology of persistent bladder inflammation in human interstitial cystitis/bladder pain syndrome. *J Urol*. 2018. <https://doi.org/10.1016/j.juro.2018.03.133>.
58. Frieling T, Cooke HJ, Wood JD. Serotonin receptors on submucous neurons in guinea pig colon. *Am J Phys*. 1991;261(6 Pt 1):G1017–23. <https://doi.org/10.1152/ajpgi.1991.261.6.G1017>.
59. Frieling T, Cooke HJ, Wood JD. Histamine receptors on submucous neurons in guinea pig colon. *Am J Phys*. 1993;264(1 Pt 1):G74–80. <https://doi.org/10.1152/ajpgi.1993.264.1.G74>.
60. Leon A, Burianni A, Dal Toso R, Fabris M, Romanello S, Aloe L, et al. Mast cells synthesize, store, and release nerve growth factor. *Proc Natl Acad Sci U S A*. 1994;91(9):3739–43. <https://doi.org/10.1073/pnas.91.9.3739>.
61. van Houwelingen AH, Kool M, de Jager SC, Redegeld FA, van Heuven-Nolsen D, Kraneveld AD, et al. Mast cell-derived TNF- $\alpha$  primes sensory nerve endings in a pulmonary hypersensitivity reaction. *J Immunol*. 2002;168(10):5297–302. <https://doi.org/10.4049/jimmunol.168.10.5297>.
62. Krumins SA, Broomfield CA. C-terminal substance P fragments elicit histamine release from a murine mast cell line. *Neuropeptides*. 1993;24(1):5–10.
63. van der Kleij HP, Ma D, Redegeld FA, Kraneveld AD, Nijkamp FP, Bienenstock J. Functional expression of neurokinin 1 receptors on mast cells induced by IL-4 and stem cell factor. *J Immunol*. 2003;171(4):2074–9. <https://doi.org/10.4049/jimmunol.171.4.2074>.
64. De Jonge F, De Laet A, Van Nassauw L, Brown JK, Miller HR, van Bogaert PP, et al. In vitro activation of murine DRG neurons by CGRP-mediated mucosal mast cell degranulation. *Am J Physiol Gastrointest Liver Physiol*. 2004;287(1):G178–91. <https://doi.org/10.1152/ajpgi.00528.2003>.
65. Dimitriadou V, Buzzi MG, Moskowitz MA, Theoharides TC. Trigeminal sensory fiber stimulation induces morphological changes reflecting secretion in rat dura mater mast cells. *Neuroscience*. 1991;44(1):97–112.
66. Ito A, Hagiya M, Oonuma J. Nerve-mast cell and smooth muscle-mast cell interaction mediated by cell adhesion molecule-1, CADM1. *J Smooth Muscle Res = Nihon Heikatsukin Gakkai kikanishi*. 2008;44(2):83–93.
67. Steers WD, Tuttle JB. Mechanisms of disease: the role of nerve growth factor in the pathophysiology of bladder disorders. *Nat Clin Pract Urol*. 2006;3(2):101–10. <https://doi.org/10.1038/ncpuro0408>.
68. Richter B, Roslind A, Hesse U, Nordling J, Johansen JS, Horn T, et al. YKL-40 and mast cells are associated with detrusor fibrosis in patients diagnosed with bladder pain syndrome/interstitial cystitis according to the 2008 criteria of the European Society for the Study of interstitial cystitis. *Histopathology*. 2010;57(3):371–83. <https://doi.org/10.1111/j.1365-2559.2010.03640.x>.
69. Anand P, Singh B, Jaggi AS, Singh N. Mast cells: an expanding pathophysiological role from allergy to other disorders. *Naunyn Schmiedeberg's Arch Pharmacol*. 2012;385(7):657–70. <https://doi.org/10.1007/s00210-012-0757-8>.
70. Homma Y, Nomiya A, Tagaya M, Oyama T, Takagaki K, Nishimatsu H, et al. Increased mRNA expression of genes involved in pronociceptive inflammatory reactions in bladder tissue of interstitial cystitis. *J Urol*. 2013;190(5):1925–31. <https://doi.org/10.1016/j.juro.2013.05.049>.
71. Tamaki M, Saito R, Ogawa O, Yoshimura N, Ueda T. Possible mechanisms inducing glomerulations in interstitial cystitis: relationship between endoscopic findings and expression of angiogenic growth factors. *J Urol*. 2004;172(3):945–8. <https://doi.org/10.1097/01.ju.0000135009.55905.cb>.
72. Kiuchi H, Tsujimura A, Takao T, Yamamoto K, Nakayama J, Miyagawa Y, et al. Increased vascular endothelial growth factor expression in patients with bladder pain syndrome/interstitial cystitis: its association with pain severity and glomerulations. *BJU Int*. 2009;104(6):826–31; discussion 31. <https://doi.org/10.1111/j.1464-410X.2009.08467.x>.
73. Jhang JF, Ho HC, Jiang YH, Lee CL, Hsu YH, Kuo HC. Electron microscopic characteristics of interstitial cystitis/bladder pain syndrome and their association with clinical condition. *PLoS One*. 2018;13(6):e0198816. <https://doi.org/10.1371/journal.pone.0198816>.
74. Saban R. Angiogenic factors, bladder neuroplasticity and interstitial cystitis—new pathobiological insights. *Transl Androl Urol*. 2015;4(5):555–62. <https://doi.org/10.3978/j.issn.2223-4683.2015.08.05>.
75. Lee JD, Lee MH. Increased expression of hypoxia-inducible factor-1 $\alpha$  and vascular endothelial growth factor associated with glomerulation formation in patients with interstitial cystitis. *Urology*. 2011;78(4):971 e11–5. <https://doi.org/10.1016/j.urology.2011.05.050>.
76. Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature*. 2000;407(6801):249–57. <https://doi.org/10.1038/35025220>.
77. •• Wennevik GE, Meijlink JM, Hanno P, Nordling J. The role of glomerulations in bladder pain syndrome: a review. *J Urol*. 2016;195(1):19–25. <https://doi.org/10.1016/j.juro.2015.06.112> **This article questioned the connection between glomerulations and IC/BPS.**
78. Chiu CD, Lee MH, Chen WC, Ho HL, Wu HC. Alexithymia and anesthetic bladder capacity in interstitial cystitis/bladder pain syndrome. *J Psychosom Res*. 2017;100:15–21. <https://doi.org/10.1016/j.jpsychores.2017.06.019>.
79. De Gucht V, Heiser W. Alexithymia and somatisation: quantitative review of the literature. *J Psychosom Res*. 2003;54(5):425–34.
80. •• Warren JW, Wessellmann U, Morozov V, Langenberg PW. Numbers and types of nonbladder syndromes as risk factors for interstitial cystitis/painful bladder syndrome. *Urology*. 2011;77(2):313–9. <https://doi.org/10.1016/j.urology.2010.08.059> **This article clearly demonstrated a significant association between somatoform disorders and IC/BPS.**
81. Chen IC, Lee MH, Lin HH, Wu SL, Chang KM, Lin HY. Somatoform disorder as a predictor of interstitial cystitis/bladder pain syndrome: evidence from a nested case-control study and a

- retrospective cohort study. *Medicine*. 2017;96(18):e6304. <https://doi.org/10.1097/MD.0000000000006304>.
82. Warren JW. Bladder pain syndrome/interstitial cystitis as a functional somatic syndrome. *J Psychosom Res*. 2014;77(6):510–5. <https://doi.org/10.1016/j.jpsychores.2014.10.003> **This article strongly implied that IC/BPS without Hunner lesions is associated with functional somatic syndrome.**
83. Bourke JH, Langford RM, White PD. The common link between functional somatic syndromes may be central sensitisation. *J Psychosom Res*. 2015;78(3):228–36. <https://doi.org/10.1016/j.jpsychores.2015.01.003>.
84. Anderson G, Berk M, Maes M. Biological phenotypes underpin the physio-somatic symptoms of somatization, depression, and chronic fatigue syndrome. *Acta Psychiatr Scand*. 2014;129(2):83–97. <https://doi.org/10.1111/acps.12182>.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.