RARE DISEASES AND LIPID METABOLISM (JAG LÓPEZ, SECTION EDITOR)

Update on Primary Hypobetalipoproteinemia

Amanda J. Hooper · John R. Burnett

Published online: 30 April 2014 © Springer Science+Business Media New York 2014

Abstract "Primary hypobetalipoproteinemia" refers to an eclectic group of inherited lipoprotein disorders characterized by low concentrations of or absence of low-density lipoprotein cholesterol and apolipoprotein B in plasma. Abetalipoproteinemia and homozygous familial hypobetalipoproteinemia, although caused by mutations in different genes, are clinically indistinguishable. A framework for the clinical follow-up and management of these two disorders has been proposed recently, focusing on monitoring of growth in children and preventing complications by providing specialized dietary advice and fat-soluble vitamin therapeutic regimens. Other recent publications on familial combined hypolipidemia suggest that although a reduction of angiopoietin-like 3 activity may improve insulin sensitivity, complete deficiency also reduces serum cholesterol efflux capacity and increases the risk of early vascular atherosclerotic changes, despite low low-density lipoprotein cholesterol levels. Specialist laboratories offer exon-by-exon sequence analysis for the molecular diagnosis of primary hypobetalipoproteinemia. In the future, massively parallel sequencing of panels of genes involved in dyslipidemia may play a greater role in the diagnosis of these conditions.

This article is part of the Topical Collection on *Rare Diseases and Lipid Metabolism*

A. J. Hooper · J. R. Burnett (⊠) Department of Clinical Biochemistry, PathWest Laboratory Medicine WA, Royal Perth Hospital, Wellington Street, GPO Box X2213, Perth, WA 6847, Australia e-mail: john.burnett@health.wa.gov.au

A. J. Hooper · J. R. Burnett School of Medicine & Pharmacology, University of Western Australia, 35 Stirling Highway, Crawley, Perth, WA 6009, Australia

A. J. Hooper

School of Pathology & Laboratory Medicine, University of Western Australia, 35 Stirling Highway, Crawley, Perth, WA 6009, Australia **Keywords** Abetalipoproteinemia · Apolipoprotein B · Chylomicron retention disease · Combined hypolipidemia · Familial hypobetalipoproteinemia · Hypobetalipoproteinemia · Low-density lipoprotein

Introduction

"Primary hypobetalipoproteinemia" (HBL) refers to an eclectic group of inherited lipoprotein disorders characterized by low concentrations (below the fifth percentile for age and sex in the population) of or absence of low-density lipoprotein (LDL) cholesterol and apolipoprotein B (apoB) in plasma, depending on the gene involved and the mode of inheritance of the condition, together with the severity of the mutation or mutations present [1].

Abetalipoproteinemia [ABL; also known as Bassen– Kornzweig syndrome; Online Mendelian Inheritance in Man (OMIM) 200100] is a very rare (less than one in one million) recessive condition characterized by the virtual absence of apoB-containing lipoproteins in plasma that typically presents early in life with the clinical manifestations of fat malabsorption, steatorrhea, and failure to thrive, progressing to ophthalmological and neurological abnormalities [2]. ABL results from mutations in both alleles of the microsomal triglyceride transfer protein (MTTP) gene (*MTTP*), a molecular chaperone critical for the formation of triglyceride-rich lipoproteins, namely, very low density lipoprotein (VLDL) and chylomicrons (CM).

Familial HBL (FHBL; OMIM 615558) is a rare (approximately one in 1,000–3,000) co-dominant condition characterized by low levels of LDL cholesterol and apoB caused by a mutation in the *APOB* gene, usually giving rise to a truncated apoB protein [3]. Affected individuals are usually asymptomatic, but are at increased risk of fatty liver disease. Inheritance of two mutations in *APOB* (compound heterozygous or homozygous FHBL) is clinically indistinguishable from ABL.

Loss-of-function mutations in *PCSK9* result in decreased LDL cholesterol and apoB levels in a gene dose-dependent manner, leading to a lifetime low risk of cardiovascular disease [4]. Heterozygous *PCSK9* nonsense mutations are found in approximately 2 % of Africans and African-Americans [5, 6], giving an estimated homozygosity of approximately one in 10,000.

CM retention disease (CMRD; OMIM 246700) is a very rare (less than one in one million) recessive condition characterized by the accumulation of lipid droplets within the enterocytes and the selective absence of apoB-48-containing particles from plasma. CMRD is caused by two mutations in *SAR1B*, the gene product of which is critical for the intracellular trafficking of CM particles [7]. Clinical manifestations of CMRD include fat malabsorption, diarrhea, abdominal distension, vomiting, and failure to thrive.

Mutations in *ANGPTL3* are associated with familial combined hypolipidemia (OMIM 605019), a recessive condition characterized by a global reduction in the levels of plasma lipoproteins [8, 9].

In this report, we provide an update on primary HBL.

Primary Hypobetalipoproteinemia

Lipoprotein disorders causing primary HBL can be classified depending on the lipid biochemical phenotype, the gene involved, and the mode of inheritance of the condition, together with the severity of the mutation or mutations present (Table 1).

Abetalipoproteinemia

In 1992, the role of MTTP was first implicated in ABL, when its activity was undetectable in intestinal biopsy specimens of individuals with ABL [10]. Mutations in the *MTTP* gene on chromosome locus 4q22-24 were subsequently described with ABL patients carrying two defective copies [11]. *MTTP* encodes an 894 amino acid protein, which forms a heterodimer with protein disulfide isomerase, thus facilitating the transfer of lipids on to nascent apoB by a shuttle mechanism [12]. Deficient MTTP activity targets apoB for degradation, preventing the secretion of triglyceride-rich lipoproteins. Mutations in *MTTP* may disrupt MTTP formation, interfere with its association with protein disulfide isomerase, or affect its ability to transfer lipids [13, 14].

ABL is associated with a variety of clinical manifestations affecting multiple organ systems. Fat malabsorption is a central feature of ABL, and is usually observed in the neonatal period with steatorrhea, vomiting, abdominal distension, and failure to thrive, and later in life, progression to atypical retinitis pigmentosa and spinocerebellar ataxia [15]. The gastrointestinal

symptoms found in ABL usually subside with age, in part, due to the avoidance of dietary fat in these patients [16]. The duodenal mucosa appears yellow on endoscopy owing to intestinal lipid accumulation [17], with histology showing normal villi with enterocytes that are distended with lipid droplets.

Acanthocytes compose up to 50 % or more of circulating erythrocytes in ABL [16] and result from either vitamin E deficiency or an altered membrane lipid composition. Other hematological abnormalities include low erythrocyte sedimentation rates, decreased red cell survival, anemia, hyperbilirubinemia, and hemolysis, and increased international normalized ratio owing to vitamin K deficiency. Liver involvement in ABL includes hepatomegaly with abnormal levels of transaminases. Liver biopsy specimens in ABL have shown marked hepatic steatosis [15], which can progress to steatohepatitis, fibrosis, and cirrhosis. The neuromuscular manifestations of ABL typically begin in the first or second decade of life, affecting both the central and the peripheral nervous system. Neurological signs which relate to vitamin E deficiency include the progressive loss of deep tendon reflexes, vibratory sense, and proprioception, muscle weakness, and eventually, a Friedrich's-like form of ataxia [18]. Ophthalmological findings tend to be variable, with many ABL patients being asymptomatic until adulthood [16]. Loss of night vision and/or color vision tends to occur early in the course of the disease. Fundoscopic examination may reveal an atypical pigmentation of the retina, which can lead to slowly enlarging scotomas that may result in blindness.

The standard treatment for ABL is dietary fat restriction with replacement of fat-soluble vitamins; however, this fails to completely control or cure this condition [1, 15, 16, 19]. A low-fat diet (less than 30 % of total calories) will eliminate steatorrhea and allows absorption of other nutrients essential for growth and development. High-dose oral vitamin E supplementation (100–300 mg/kg/day) is recommended to halt the progression of the neurological disease; however, serum levels do not fully normalize [15, 16]. Patients need to be followed up regularly for evaluation of symptoms and complications, and to monitor them for compliance with therapy [20••].

Supplementation with a combination of high-dose vitamins E and A is effective in reducing retinal degeneration [21]. Patients treated with high-dose vitamin E from the age of 16 months do not develop neurological or retinal features, and progression is halted or sometimes even reversed in older patients who already show symptoms of neurological dys-function [22]. Although serum vitamin E is usually undetectable in untreated ABL, supplementation results in trace concentrations, with normal levels in adipose tissue [23]. Erythrocyte and platelet vitamin E have also been used to assess tissue vitamin E status [24]. Although vitamin D and vitamin K deficiencies are inconsistent findings in ABL, oral replacement should be considered, along with other supplementary nutrients such as iron and folate if required.

Table 1 Lipoprotein disorders causing primary hypobetalipoproteinemia

Lipoprotein disorder	Gene	Inheritance	Biochemical phenotype	Clinical phenotype
Abetalipoproteinemia	MTTP	Recessive	Absence of LDL and chylomicrons, low levels of triglycerides, very low levels of vitamin E	Variable; includes failure to thrive, steatorrhea, and progressive neurological and ophthalmological abnormalities
Familial hypobetalipoproteinemia	APOB	Co-dominant	Heterozygous: LDL cholesterol levels less than 30 % of levels normal for age and sex. Homozygous: absence of or very low levels of LDL cholesterol, low levels of triglycerides, very low levels of vitamin E	Heterozygous: generally asymptomatic, may include fatty liver. Homozygous: indistinguishable from abetalipoproteinemia
Familial hypobetalipoproteinemia	PCSK9	Co-dominant	Heterozygous: approximately 40 % reduction in LDL levels. Homozygous: very low LDL cholesterol levels	None
Chylomicron retention disease	SAR1B	Recessive	Absence of chylomicrons, LDL cholesterol levels less than 50 % of levels normal for age and sex	Variable; includes failure to thrive, steatorrhea, and progressive neurological abnormalities
Familial combined hypolipidemia	ANGPTL3	Recessive	Reduced levels of all plasma lipoproteins	None

LDL low-density lipoprotein

A framework for the clinical follow-up and management of ABL (and compound heterozygous and homozygous FHBL) has been proposed recently, focusing on monitoring of growth in children and detecting and preventing complications in all ABL subjects by providing specialized dietary advice and fatsoluble vitamin therapeutic regimens [20••].

Familial Hypobetalipoproteinemia

Mutations in the *APOB* gene on chromosome locus 2p23-24 either abolish or interfere with the translation of full-length apoB and cause FHBL [25–27]. Most mutations are nucleotide substitutions and deletions in exon 26. The resulting apoB truncations have traditionally been named according to a centile system. In addition, R463W and several other missense mutations in the N-terminal $\beta \alpha_1$ domain of apoB have also been described [28–30]. R463W causes impaired secretion of VLDL by defective intracellular transport and enhanced binding of the mutant protein to MTTP, leading to FHBL [28].

In patients with FHBL, truncated forms of apoB are produced at lower rates than apoB-100 and have reduced lipid content [31]. As truncated apoBs shorter than apoB-30 are not detectable in plasma, the corresponding length appears to be the minimum length of apoB that is required for lipoprotein assembly. Truncations shorter than this are unable to be sufficiently lipidated and are targeted for intracellular degradation [32].

FHBL can also be caused by mutations in *PCSK9*, which encodes a protease that binds to the LDL receptor and targets it for lysosomal degradation within hepatocytes [33]. Nonsense and other loss-of-function *PCSK9* missense mutations increase the number of LDL receptors on the cell surface, thereby reducing both circulating LDL cholesterol

concentrations and coronary heart disease risk. Heterozygous carriers of the *PCSK9* nonsense mutations Y142X and C679X had LDL cholesterol concentrations reduced by 28 %, with a corresponding reduction in coronary heart disease risk of 88 %, compared with noncarriers [4]. A compound heterozygote and a homozygote for *PCSK9* nonsense mutations have been described, each healthy and with a very low LDL cholesterol concentration of approximately 0.4 mmol/L [6, 34]. Human kinetic studies have confirmed that loss-of-function *PCSK9* mutations increase the fractional catabolic rate of LDL [35].

Although patients with heterozygous *APOB*-linked FHBL are often asymptomatic, many develop fatty liver [36, 37]. FHBL might represent a longevity syndrome due to a lower lifetime exposure to atherogenic apoB-containing lipoproteins. However, only the surrogate markers carotid intimamedia thickness (CIMT) and distal common carotid arterial wall stiffness have been used to demonstrate the cardioprotective effects of FHBL in humans [38].

The long-term consequences of the increased levels of liver transaminases and hepatic steatosis seen in heterozygous FHBL are unknown. FHBL heterozygotes have threefold to fivefold greater liver fat content compared with control subjects [37, 39, 40]. Regular biochemical monitoring and liver imaging is recommended, given the potential to progress to cirrhosis, particularly in the presence of known risk factors, such as alcohol, caloric excess, and liver injury [26, 41–43].

The clinical and biochemical features of homozygous (and compound heterozygous) FHBL are, in general, indistinguishable from those of ABL. Dietary fats should be restricted to prevent steatorrhea, and long-term high-dose vitamin E and vitamin A supplementation should be given to prevent or at least slow the progression of neuromuscular and retinal degenerative disease [16, 21].

Chylomicron Retention Disease

Mutations in *SAR1B*, a member of the Sar1-ADP-ribosylation factor family of small GTPases that control the intracellular trafficking of proteins, are the cause of CMRD [44]. SAR1B is needed for the fusion of the intestine-specific pre-CM transport vesicle to the Golgi apparatus, allowing transport of CM through the cellular secretory pathways [45]. Mutations in *SAR1B* result in the inability to secrete CM, leading to the accumulation of lipid droplets within the enterocytes.

CMRD presents shortly after birth with diarrhea, fat malabsorption, and failure to thrive, with vomiting and abdominal distension often present [7, 46]. Acanthocytosis is rare and may be transient. Hepatomegaly and hepatic steatosis may develop in some patients, along with fat-soluble vitamin deficiencies and their manifestations. In contrast to ABL and FHBL, cirrhosis has not been reported in CMRD. The duodenal mucosa appears white on endoscopy, with vacuolization of enterocytes in intestinal villi of normal structure present on histology, findings similar to those in ABL.

Patients with CMRD improve within days or weeks on commencement of a low-fat diet of polyunsaturated fatty acids [7]. No relationship has been found between liver transaminases, hepatomegaly, and hepatic steatosis. Neurological features include hyporeflexia and loss of proprioception in adolescents through to ataxia, myopathy, and sensory neuropathy in adults.

There are no specific recommendations for the follow-up or treatment of CMRD, with therapeutic regimens currently based on those recommended for ABL [20••].

Familial Combined Hypolipidemia

Homozygosity (or compound heterozygosity) for mutations in *ANGPTL3*, the gene encoding angiopoietin-like 3 (ANGPTL3), are associated with combined hypolipidemia, which is characterized by extremely low plasma levels of LDL cholesterol, high-density lipoprotein (HDL) cholesterol, and triglyceride [9]. The function of ANGPTL3 appears to be the reversible inhibition of lipase activity [47, 48]. The disruption of ANGPTL3 production increases lipolysis, enhancing clearance of lipoproteins and decreasing circulating lipid concentrations.

The Italian town of Campodimele has been the focus of studies into familial combined hypolipidemia. In 1991, a three-generation family was described with a dominant form of HBL that was not due to a mutation in *APOB* [49]. The *ANGPTL3* S17X mutation was subsequently identified in the proband of this family and in an additional eight local families [50]. The prevalence of *ANGPTL3* variants in the population was estimated to be 9.4 % [50].

A pooled analysis of *ANGPTL3* mutation carriers, including 14 homozygotes, eight compound heterozygotes, and 93 heterozygotes, confirmed that heterozygotes as well as homozygotes had a significant reduction in the levels of all plasma lipoproteins compared with 402 controls [51]. Consistent with earlier reports showing that there was no difference in the prevalence of elevated plasma concentrations of liver enzymes [50], the prevalence of fatty liver was not different among groups [51]. However, diabetes and cardiovascular disease were absent in homozygotes, raising the possibility that absence of ANGPTL3 is protective for these conditions.

Recently it was reported that *ANGPTL3* S17X homozygotes showed significantly lower plasma insulin concentrations and a trend towards lower plasma glucose concentrations [52•]. The absence of ANGPTL3 results in an increase in lipoprotein lipase activity and reduced plasma free fatty acid levels, which may improve tissue insulin action.

Surprisingly, although partial ANGPTL3 deficiency (as seen in heterozygous mutation carriers) is not associated with vascular changes, homozygosity leading to complete ANGPTL3 deficiency may be associated with an increased risk of developing early vascular atherosclerotic changes despite low plasma LDL cholesterol levels [53••]. Homozygotes showed an increased CIMT, with a 0.19 mm higher average CIMT and a 0.54 mm higher maximum CIMT, along with a trend towards lower brachial flow-mediated dilatation. This may relate to impaired HDL function, as homozygotes also showed a reduction in serum cholesterol efflux capacity.

The prevalence of *ANGPTL3* mutations giving rise to a combined hypolipidemia phenotype in subjects with severe primary HBL is about 10 % [54]. Of subjects with total cholesterol concentration below the second percentile, those with HDL cholesterol concentration below the second decile may be carrying *ANGPTL3* mutations, whereas those with higher HDL cholesterol concentrations may be carrying *APOB* mutations [54].

An Approach to the Patient with Primary Hypobetalipoproteinemia

The usual biochemical trigger for the investigation of suspected primary HBL is the finding of marked hypocholesterolemia with plasma LDL cholesterol and apoB concentrations below the fifth percentile for age and sex [1]. A personal and family history should be taken, and a physical examination should be conducted. Secondary causes of HBL, such as severe chronic liver disease, chronic pancreatitis, cystic fibrosis, end-stage renal disease, hyperthyroidism, cachexia, and malabsorption should be excluded, and clinical manifestations, such as fat malabsorption, growth failure, fatsoluble vitamin deficiency, fatty liver disease, and neurological and ophthalmological dysfunction should be sought.

Patients with ABL (and compound heterozygous and homozygous FHBL) will have very low plasma total cholesterol and generally low triglyceride concentrations. LDL cholesterol, when measured by direct methods, and apoB, will be absent or their concentrations will be very low. Vitamin E levels will also be very low, and acanthocytosis may be observed on peripheral blood smear.

Patients with heterozygous FHBL typically have plasma LDL cholesterol and apoB concentrations that are one quarter to one third of normal. The reasons for these lower-thanexpected levels may include decreased hepatic secretion of the apoB-containing lipoproteins, or the upregulation of the LDL receptor, resulting in an enhanced clearance rate for VLDL and LDL particles produced by the normal allele [31].

Subjects who carry a single *MTTP* mutation may have normal plasma lipid levels or may have LDL cholesterol and apoB concentrations similar to those seen in heterozygous FHBL.

In CMRD, total cholesterol, LDL cholesterol, and HDL cholesterol concentrations are low, but triglyceride levels are generally normal. The low plasma LDL cholesterol and HDL cholesterol concentrations are a consequence of low rates of apoB-100 and apolipoprotein A-I production [55]. An increased plasma creatine kinase concentration of up to five times the normal level may be seen from infancy, along with deficiencies in fat-soluble vitamins [7].

If concomitant reductions in triglyceride and HDL cholesterol concentrations are observed, this is suggestive of familial combined hypolipidemia.

Molecular Tests

Sequencing of the HBL genes *MTTP*, *APOB*, *SAR1B*, *PCSK9*, and *ANGPTL3* is available in specialist laboratories. These molecular assays are usually designed to target exonic regions as well as at least 20 base pairs of flanking intronic sequence in order to capture any potential splice site mutations. Where two mutations are identified, testing of the patient's parents is recommended to confirm that the mutations originate from two different chromosomes.

Western blotting can be used to detect truncated apoB species that are more than 30 % of the size of the full-length protein. DNA sequencing of the region in *APOB* where the mutation is estimated to occur can then be performed. However, as truncated apoB species shorter than apoB-30 are not detectable in plasma, in the situation of a negative Western blot result, then sequencing of the first 30 % of the *APOB* gene (exons 1–25) should be performed.

In patients with ABL where the inheritance pattern is unclear or *MTTP* mutations cannot be identified, then the *APOB* gene should also be sequenced, given the clinical and biochemical similarities with compound heterozygous and homozygous FHBL. Likewise, in patients with homozygous FHBL, the *MTTP* gene could be sequenced in the event where *APOB* mutations cannot be found. Alternatively, high-throughput sequencing technology is emerging as a means for screening multiple genes for mutations. This may prove more costeffective than traditional Sanger sequencing, particularly where multiple genes may need to be sequenced and when the *APOB* gene is involved, which needs over 40 primer sets to cover the whole coding region. Recently, a next-generation resequencing approach for the diagnosis of monogenic dyslipidemias has been described; this targets a customized panel of 73 genes, including those associated with HBL, and has the potential to diagnose patients rapidly and at much lower cost than traditional Sanger or whole-exome sequencing [56••].

Conclusion

In this report, we have provided an update on primary HBL, an eclectic group of inherited lipoprotein disorders characterized by low concentrations of or absence of LDL cholesterol and apoB in plasma, depending on the gene involved and the mode of inheritance of the condition, together with the severity of the mutation or mutations present. Some of these lipoprotein disorders, such as ABL (and compound heterozygous and homozygous FHBL), are associated with clinical manifestations of fat malabsorption, growth failure, fat-soluble vitamin deficiency, fatty liver disease, and neuro-ophthalmological dysfunction, whereas others, such as heterozygous FHBL, may be asymptomatic. Molecular testing of the MTTP, APOB, SAR1B, PCSK9, and ANGPTL3 genes is available in specialist laboratories. A framework for the clinical follow-up and management of ABL (and compound heterozygous and homozygous FHBL) has been proposed recently, focusing on monitoring of growth in children and detecting and preventing complications in all ABL subjects by providing specialized dietary advice and fat-soluble vitamin therapeutic regimens.

Acknowledgments This work was supported by National Health and Medical Research Council Project Grant 1010133 (to Amanda J. Hooper and John R. Burnett) and a Practitioner Fellowship from the Royal Perth Hospital Medical Research Foundation (to John R. Burnett).

Conflict of Interest Amanda J. Hooper and John R. Burnett declare that they have no conflict of interest.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
 - Hooper AJ, van Bockxmeer FM, Burnett JR. Monogenic hypocholesterolaemic lipid disorders and apolipoprotein B metabolism. Crit Rev Clin Lab Sci. 2005;42:515–45.

- Burnett JR, Bell DA, Hooper AJ, Hegele RA. Clinical utility gene card for: abetalipoproteinaemia. Eur J Hum Genet. 2012. doi:10. 1038/ejhg.2012.30.
- Burnett JR, Bell DA, Hooper AJ, Hegele RA. Clinical utility gene card for: familial hypobetalipoproteinaemia (APOB). Eur J Hum Genet. 2012. doi:10.1038/ejhg.2012.85.
- Cohen JC, Boerwinkle E, Mosley Jr TH, Hobbs HH. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. N Engl J Med. 2006;354:1264–72.
- Cohen J, Pertsemlidis A, Kotowski IK, Graham R, Garcia CK, Hobbs HH. Low LDL cholesterol in individuals of African descent resulting from frequent nonsense mutations in PCSK9. Nat Genet. 2005;37:161–5.
- Hooper AJ, Marais AD, Tanyanyiwa DM, Burnett JR. The C679X mutation in PCSK9 is present and lowers blood cholesterol in a Southern African population. Atherosclerosis. 2007;193:445–8.
- 7. Peretti N, Sassolas A, Roy CC, et al. Guidelines for the diagnosis and management of chylomicron retention disease based on a review of the literature and the experience of two centers. Orphanet J Rare Dis. 2010;5:24.
- Hooper AJ, Burnett JR. Recent developments in the genetics of LDL deficiency. Curr Opin Lipidol. 2013;24:111–5.
- Musunuru K, Pirruccello JP, Do R, et al. Exome sequencing, ANGPTL3 mutations, and familial combined hypolipidemia. N Engl J Med. 2010;363:2220–7.
- Wetterau JR, Aggerbeck LP, Bouma ME, et al. Absence of microsomal triglyceride transfer protein in individuals with abetalipoproteinemia. Science. 1992;258:999–1001.
- Shoulders CC, Brett DJ, Bayliss JD, et al. Abetalipoproteinemia is caused by defects of the gene encoding the 97 kDa subunit of a microsomal triglyceride transfer protein. Hum Mol Genet. 1993;2: 2109–16.
- 12. Hussain MM, Rava P, Walsh M, Rana M, Iqbal J. Multiple functions of microsomal triglyceride transfer protein. Nutr Metab (Lond). 2012;9:14.
- 13. Ohashi K, Ishibashi S, Osuga J, et al. Novel mutations in the microsomal triglyceride transfer protein gene causing abetalipoproteinemia. J Lipid Res. 2000;41:1199–204.
- 14. Rehberg EF, Samson-Bouma ME, Kienzle B, et al. A novel abetalipoproteinemia genotype. Identification of a missense mutation in the 97-kDa subunit of the microsomal triglyceride transfer protein that prevents complex formation with protein disulfide isomerase. J Biol Chem. 1996;271:29945–52.
- Berriot-Varoqueaux N, Aggerbeck LP, Samson-Bouma M, Wetterau JR. The role of the microsomal triglyceride transfer protein in abetalipoproteinemia. Annu Rev Nutr. 2000;20:663–97.
- Kane JP, Havel RJ. Disorders of the biogenesis and secretion of lipoproteins containing the B apolipoproteins. In: Scriver CR, Beaudet AL, Sly WS, Scriver CR, Beaudet AL, Sly WS, editors. The metabolic and molecular bases of inherited disease. New York: McGraw-Hill; 2001. p. 2717–52.
- Delpre G, Kadish U, Glantz I, Avidor I. Endoscopic assessment in abetalipoproteinemia (Bassen-Kornzweig-syndrome). Endoscopy. 1978;10:59–62.
- Tanyel MC, Mancano LD. Neurologic findings in vitamin E deficiency. Am Fam Physician. 1997;55:197–201.
- Zamel R, Khan R, Pollex RL, Hegele RA. Abetalipoproteinemia: two case reports and literature review. Orphanet J Rare Dis. 2008;3: 19.
- 20.•• Lee J, Hegele RA. Abetalipoproteinemia and homozygous hypobetalipoproteinemia: a framework for diagnosis and management. J Inherit Metab Dis. 2014. doi:10.1007/s10545-013-9665-4. *This article discusses the diagnosis, assessment, treatment, and follow-up of ABL and homozygous FHBL.*
- 21. Chowers I, Banin E, Merin S, Cooper M, Granot E. Long-term assessment of combined vitamin A and E treatment for the

prevention of retinal degeneration in abetalipoproteinaemia and hypobetalipoproteinaemia patients. Eye. 2001;15:525–30.

- 22. Muller DP. Vitamin E, and neurological function. Mol Nutr Food Res. 2010;54:710–8.
- Kayden HJ, Hatam LJ, Traber MG. The measurement of nanograms of tocopherol from needle aspiration biopsies of adipose tissue: normal and abetalipoproteinemic subjects. J Lipid Res. 1983;24:652–6.
- Clarke MW, Hooper AJ, Headlam HA, Wu JH, Croft KD, Burnett JR. Assessment of tocopherol metabolism and oxidative stress in familial hypobetalipoproteinemia. Clin Chem. 2006;52:1339–45.
- 25. Schonfeld G, Lin X, Yue P. Familial hypobetalipoproteinemia: genetics and metabolism. Cell Mol Life Sci. 2005;62:1372–8.
- Tarugi P, Averna M. Hypobetalipoproteinemia: genetics, biochemistry, and clinical spectrum. Adv Clin Chem. 2011;54:81–107.
- Whitfield AJ, Barrett PHR, van Bockxmeer FM, Burnett JR. Lipid disorders and mutations in the APOB gene. Clin Chem. 2004;50: 1725–32.
- Burnett JR, Shan J, Miskie BA, et al. A novel nontruncating APOB gene mutation, R463W, causes familial hypobetalipoproteinemia. J Biol Chem. 2003;278:13442–52.
- Burnett JR, Zhong S, Jiang ZG, et al. Missense mutations in APOB within the betaalpha1 domain of human APOB-100 result in impaired secretion of apoB and apoB-containing lipoproteins in familial hypobetalipoproteinemia. J Biol Chem. 2007;282:24270–83.
- Zhong S, Magnolo AL, Sundaram M, et al. Nonsynonymous mutations within APOB in human familial hypobetalipoproteinemia evidence for feedback inhibition of lipogenesis and postendoplasmic reticulum degradation of apolipoprotein B. J Biol Chem. 2010;285:6453–64.
- Elias N, Patterson BW, Schonfeld G. Decreased production rates of VLDL triglycerides and apoB-100 in subjects heterozygous for familial hypobetalipoproteinemia. Arterioscler Thromb Vasc Biol. 1999;19:2714–21.
- 32. Yao ZM, Blackhart BD, Linton MF, Taylor SM, Young SG, McCarthy BJ. Expression of carboxyl-terminally truncated forms of human apolipoprotein B in rat hepatoma cells. Evidence that the length of apolipoprotein B has a major effect on the buoyant density of the secreted lipoproteins. J Biol Chem. 1991;266:3300–8.
- Lambert G, Sjouke B, Choque B, Kastelein JJ, Hovingh GK. The PCSK9 decade. J Lipid Res. 2012;53:2515–24.
- 34. Zhao Z, Tuakli-Wosornu Y, Lagace TA, et al. Molecular characterization of loss-of-function mutations in PCSK9 and identification of a compound heterozygote. Am J Hum Genet. 2006;79:514–23.
- Cariou B, Ouguerram K, Zair Y, et al. PCSK9 dominant negative mutant results in increased LDL catabolic rate and familial hypobetalipoproteinemia. Arterioscler Thromb Vasc Biol. 2009;29:2191–7.
- Ogata H, Akagi K, Baba M, et al. Fatty liver in a case with heterozygous familial hypobetalipoproteinemia. Am J Gastroenterol. 1997;92:339–42.
- Schonfeld G, Patterson BW, Yablonskiy DA, et al. Fatty liver in familial hypobetalipoproteinemia: triglyceride assembly into VLDL particles is affected by the extent of hepatic steatosis. J Lipid Res. 2003;44:470–8.
- Sankatsing RR, Fouchier SW, de Haan S, et al. Hepatic and cardiovascular consequences of familial hypobetalipoproteinemia. Arterioscler Thromb Vasc Biol. 2005;25:1979–84.
- Tanoli T, Yue P, Yablonskiy D, Schonfeld G. Fatty liver in familial hypobetalipoproteinemia: roles of the APOB defects, intraabdominal adipose tissue, and insulin sensitivity. J Lipid Res. 2004;45:941–7.
- 40. Visser ME, Lammers NM, Nederveen AJ, et al. Hepatic steatosis does not cause insulin resistance in people with familial hypobetalipoproteinaemia. Diabetologia. 2011;54:2113–21.

- 41. Hooper AJ, Adams LA, Burnett JR. Genetic determinants of hepatic steatosis in man. J Lipid Res. 2011;52:593–617.
- 42. Lonardo A, Tarugi P, Ballarini G, Bagni A. Familial heterozygous hypobetalipoproteinemia, extrahepatic primary malignancy, and hepatocellular carcinoma. Dig Dis Sci. 1998;43:2489–92.
- 43. Tarugi P, Lonardo A, Ballarini G, et al. A study of fatty liver disease and plasma lipoproteins in a kindred with familial hypobetalipoproteinemia due to a novel truncated form of apolipoprotein B (APO B-54.5). J Hepatol. 2000;33:361–70.
- 44. Jones B, Jones EL, Bonney SA, et al. Mutations in a Sar1 GTPase of COPII vesicles are associated with lipid absorption disorders. Nat Genet. 2003;34:29–31.
- Siddiqi SA, Gorelick FS, Mahan JT, Mansbach 2nd CM. COPII proteins are required for Golgi fusion but not for endoplasmic reticulum budding of the pre-chylomicron transport vesicle. J Cell Sci. 2003;116:415–27.
- Peretti N, Roy CC, Sassolas A, et al. Chylomicron retention disease: a long term study of two cohorts. Mol Genet Metab. 2009;97: 136–42.
- Nakajima K, Kobayashi J, Mabuchi H, et al. Association of angiopoietin-like protein 3 with hepatic triglyceride lipase and lipoprotein lipase activities in human plasma. Ann Clin Biochem. 2010;47:423–31.
- Shan L, Yu XC, Liu Z, et al. The angiopoietin-like proteins ANGPTL3 and ANGPTL4 inhibit lipoprotein lipase activity through distinct mechanisms. J Biol Chem. 2009;284:1419–24.
- Fazio S, Sidoli A, Vivenzio A, et al. A form of familial hypobetalipoproteinaemia not due to a mutation in the apolipoprotein B gene. J Intern Med. 1991;229:41–7.
- Minicocci I, Montali A, Robciuc MR, et al. Mutations in the ANGPTL3 gene and familial combined hypolipidemia: a clinical

and biochemical characterization. J Clin Endocrinol Metab. 2012;97:E1266–75.

- Minicocci I, Santini S, Cantisani V, et al. Clinical characteristics and plasma lipids in subjects with familial combined hypolipidemia: a pooled analysis. J Lipid Res. 2013;54:3481–90.
- 52.• Robciuc MR, Maranghi M, Lahikainen A, et al. Angptl3 deficiency is associated with increased insulin sensitivity, lipoprotein lipase activity, and decreased serum free fatty acids. Arterioscler Thromb Vasc Biol. 2013;33:1706–13. *This article shows that although partial deficiency of ANGPTL3 did not affect lipase activity, complete deficiency decreased the levels of free fatty acids and improved insulin sensitivity.*
- 53.•• Minicocci I, Cantisani V, Poggiogalle E, et al. Functional and morphological vascular changes in subjects with familial combined hypolipidemia: an exploratory analysis. Int J Cardiol. 2013;168: 4375–8. Despite an approximately 50% reduction in the concentration of LDL cholesterol, ANGPTL3 homozygotes had increased CIMT.
- 54. Noto D, Cefalu AB, Valenti V, et al. Prevalence of ANGPTL3 and APOB gene mutations in subjects with combined hypolipidemia. Arterioscler Thromb Vasc Biol. 2012;32:805–9.
- Ouguerram K, Zair Y, Kasbi-Chadli F, et al. Low rate of production of apolipoproteins B100 and AI in 2 patients with Anderson disease (chylomicron retention disease). Arterioscler Thromb Vasc Biol. 2012;32:1520–5.
- 56.•• Johansen CT, Dube JB, Loyzer MN, et al. LipidSeq: a nextgeneration clinical resequencing panel for monogenic dyslipidemias. J Lipid Res. 2014;55:765–72. This article describes the design and performance of a targeted resequencing panel which has potential to aid in molecular diagnosis of dyslipidemias, including primary HBL.