

# T-cell immunity against cytomegalovirus in HIV infection and aging: relationships with inflammation, immune activation, and frailty

Juliette Tavenier<sup>1</sup> · Joseph B. Margolick<sup>2</sup> · Sean X. Leng<sup>3</sup>

Received: 5 February 2019 / Accepted: 6 March 2019 / Published online: 21 March 2019 © Springer-Verlag GmbH Germany, part of Springer Nature 2019

#### Abstract

Both aging and treated human immunodeficiency virus (HIV) infection are characterized by low-level chronic inflammation and immune activation which contribute to the development of age-related diseases, frailty, and early mortality. Chronic cytomegalovirus (CMV) infection is highly prevalent in older adults and HIV-infected populations. A number of studies have shown that CMV induces broad and strong T-cell responses in CMV-seropositive older adults and HIV-infected individuals. CMV infection rarely develops into clinical disease in immunocompetent individuals. However, a large body of literature has shown adverse effects of chronic CMV infection on the health and longevity of these populations. It has been hypothesized that chronic CMV infection may be a driver of chronic inflammation and immune activation, and may further contribute to the development of frailty. Thus, there is a need to better understand the extent of the impact of chronic CMV infection on T-cell immunity and health in aging and HIV infection. In this review, we will address important considerations and challenges in the assessment of chronic CMV infection and CMV-specific T-cell responses. We will then review recent data on relationships between T-cell responses to CMV and levels of inflammatory markers and immune activation, as well as the onset of frailty.

Keywords Cytomegalovirus · Frailty · CLIP · Immune activation · HIV infection · Aging

## Introduction

Aging is accompanied by a chronic low-grade inflammatory phenotype (CLIP) marked by elevated levels of circulating inflammatory mediators including C-reactive protein (CRP)

Edited by: Matthias J. Reddehase.

This article is part of the Special Issue on Immunological Imprinting during Chronic Viral Infection.

Sean X. Leng sleng1@jhmi.edu

- Present Address: Clinical Research Centre, Copenhagen University Hospital Hvidovre, Kettegaard Alle 30, 2650 Hvidovre, Denmark
- <sup>2</sup> Present Address: Department of Molecular Microbiology and Immunology, Johns Hopkins Bloomberg School of Public Health, 615 N Wolfe Street, Room E5153, Baltimore, MD 21205-7535, USA
- <sup>3</sup> Present Address: Division of Geriatric Medicine and Gerontology, Department of Medicine, Johns Hopkins University School of Medicine, 5501 Hopkins Bayview Circle-Room 1A.38A, Baltimore, MD 21224, USA

and interleukin 6 (IL-6) [1], as well as chronic immune activation, both of which are believed to contribute significantly to most age-related diseases such as cardiovascular and neurodegenerative disease, frailty, and cancer [2-4]. While it is believed that HIV-infected (HIV+) individuals experience accelerated aging of their immune system, they differ from the age-related immune changes, or immunosenescence, in HIV-uninfected (HIV-) older adults. For example, HIV infection results in the depletion of CD4 T cells, and likely massive destruction of the gut barrier. Nevertheless, in parallel with what is observed in the geriatric population, well-treated HIV+ persons exhibit elevated levels of inflammatory mediators and chronic immune activation despite effective viral suppression [5, 6]. Although chronic inflammation in treated HIV infection is likely the result of residual HIV replication, co-infections, or increased microbial translocation due to damage to the gut mucosa, markers of inflammation and immune activation that are elevated as well as their associations with comorbidities are very much alike between HIV- and treated HIV+ aging populations [7]. Although difficult to quantify, HIV+ individuals manifest earlier onset and increased prevalence of age-related comorbidities and frailty compared to HIV- aging population [7]. Frailty is a state of increased vulnerability resulting from decline in physiological reserve and dysregulation of multiple organ systems. CLIP and chronic immune activation are thought to contribute to frailty in the elderly and in HIV+ individuals [2, 8]. The etiology of CLIP and immune activation in aging is unknown, but chronic cytomegalovirus (CMV) infection is common in the older adult and HIV+ populations and is suspected to contribute to chronic inflammation and immune activation.

CMV is a beta-herpesvirus, and is the largest virus known to cause human disease. Following primary infection, CMV can maintain a state of latency in myeloid progenitor cells and monocytes, and can be reactivated following inflammatory immune response or critical illness such as sepsis, differentiation of infected cells, or immunosuppression, although the exact mechanisms and conditions that trigger reactivation remain to be further elucidated. CMV seroprevalence increases with age and reaches 90% in those aged 80 years and over and is almost universal in HIV+ populations [9–11]. CMV is unique in its ability to drive clonal expansion of CMV-specific T-cells even in the absence of overt active infection. This results in skewing of the T-cell repertoire towards CMV antigens, often comprising more than 10% of T-cells [11–20]. While CMV infection rarely leads to clinical symptoms in immunocompetent persons and is even thought to have protective effects in youth [21], a large body of literature has associated CMV infection with adverse outcomes in health and longevity [22-26]. However, these observations are for the most part derived from cross-sectional studies and insight into the mechanisms and causality behind these associations are still lacking, and longitudinal and mechanistic studies to explore causality are needed. In people with advanced HIV infection, inhibition of CMV replication using valganciclovir significantly decreased the percentage of activated (CD38<sup>+</sup>HLADR<sup>+</sup>) CD8 T-cells, supporting the idea of a link between CMV infection and immune activation [27]. However, the relationships of T-cell responses to CMV with CLIP, immune activation and onset of frailty in the HIV+ aging population have just begun to be elucidated. In this article, we will first address important considerations and challenges the assessment of chronic CMV infection and CMV T-cell responses. We will then review recent data on correlations between T-cell responses to CMV and levels of inflammatory markers and immune activation, as well as the onset of frailty.

# Challenges in assessing chronic CMV infection

CMV is a complex virus whose infection presents a diagnostic challenge. CMV infection is considered a smoldering infection as the virus establishes latency, but may

periodically reactivate in some cells or tissues, resulting in low-level viral replication. Traditionally, the most common method is the measurement of CMV-reactive antibodies, anti-CMV IgG for chronic infection and IgM primary or re-infection. Based on anti-CMV IgG serology, individuals are routinely classified as CMV-seropositive or negative, and some may use absolute anti-CMV IgG titers to assess CMV burden. Therefore, anti-CMV IgG serology is the current diagnostic paradigm for chronic CMV infection both in older adults and in HIV+ aging individuals. However, anti-CMV IgG serology is a crude measure that merely indicates prior exposure to the virus. It does not allow to assess the degree or frequency of reactivation and low-level viral replication. A longitudinal analysis of a subset of participants from the Women's Health and Aging Studies (WHAS) II showed essentially no change in absolute anti-CMV IgG titers over a 12-year follow-up period, indicating limited utility, if any, of this measure for monitoring of chronic CMV infection [18]. Other diagnostic evaluation tools include different PCR methods including nested PCR and droplet digital PCR (ddPCR) for the detection of cell-associated CMV DNA which may allow more accurate evaluation of the current state of CMV infection and reactivation [28]. We have shown that the presence of CMV UL123 and UL93 DNA in monocyte-enriched peripheral blood mononuclear cells (PBMCs) rather than anti-CMV IgG titers was associated with high frequencies of CMV pp65-specific CD8 T-cells in older adults [17]. This may explain why some studies reported no significant association between anti-CMV IgG titers and adverse health outcomes [28-30]. Although highly sensitive, nested PCR is a qualitative assay. On the other hand, ddPCR allows accurate quantification of low numbers of target DNA copies. One study reports that CMV prevalence and viral load in the peripheral monocytes increases with age in a cohort of healthy donors [31]. Future studies should validate these novel diagnostic tools and further investigate the relationship between chronic CMV infection as defined by these novel methods and T-cell responses to CMV.

Assessing T-cell responses to CMV has also proven to be a complex matter. A vast majority of studies assess T-cell responses to CMV by only measuring responses to the CMV phosphoprotein 65, pp65 (encoded by UL83) and immediate early-1, IE1 (UL123) epitopes through tetramer, pentamer or dextramer analysis, as it was assumed that these represented the dominant responses [12–15, 23, 32–35]. However, using ribosome profiling and transcript analysis, Stern-Ginossar et al. observed over 700 open reading frames (ORFs) in human CMV genome that are translated to protein in CMVinfected fibroblasts [36]. Although many of these proteins may only be short lived and non-functional, the antigenic potential of CMV is likely much greater than previously anticipated. In a study of 33 healthy CMV-seropositive donors, peptide pools from 213 CMV ORFs (a total of over 13,000 peptides) were tested for immunogenicity, and 151 of these ORFs were recognized by either CD4 T-cells (44 ORFs), CD8 T-cells (26 ORFs), or both (81 ORFs) [19]. The ORFs recognized by CD4 and CD8 T-cells were different, but 8 of the 15 most recognized ORFs were common to both CD4 and CD8 T-cells including UL83 and UL123. In addition to the broad range of recognized CMV epitopes, the T-cell responses to CMV are highly heterogeneous. The number of CMV ORFs recognized by T-cells from healthy donors varied greatly from 5 to 55 ORFs. Thus, to accurately assess the overall CMV-specific T-cell responses, one or a few ORFs are not sufficient, at least the 19 top ORFs should be used to obtain an acceptable approximation of the overall T-cell response to CMV [19]. In 12 virologically suppressed HIV+ and 10 HIV- CMV-seropositive men from the Multicenter AIDS Cohort Study (MACS) of men who have sex with men (MSM), we evaluated CD4 and CD8 T-cell responses to the previously described 19 most recognized ORFs [37]. We found that only < 12% and < 40%of total CMV-specific CD4 and CD8 T-cells, respectively, responded to pp65 or IE1 peptide pools, and there was no difference between HIV+ and HIV- individuals. This supports the notion that data restricted to these two epitopes do not represent the overall CMV-specific T-cell immunity.

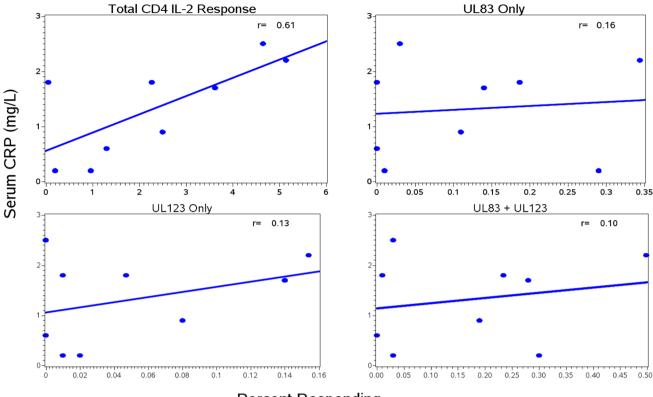
We and others have also demonstrated the heterogeneity in the magnitude and breadth of T-cell responses to CMV within older adults and within HIV+ individuals [37-39]. In several studies, although the mean number of CMV-specific T-cells was elevated in CMV-seropositive older age group, there was a large variation between individuals. Many of the individuals in the older age group had similar number of CMV-specific T-cells as seronegative or young individuals [20, 23, 28]. It remains unclear whether broader and greater T-cell responses are required for protection against CMV. Given the rarity of CMV-related disease in immunocompetent individuals, it is possible that very low and restricted T-cell responses are sufficient to protect from CMV disease. This raises the question of whether broader and greater T-cell responses are excessive and may prove to be detrimental by leading to immunosenescence, increased chronic inflammation and immune activation, and thereby contributing to frailty and other age-related chronic conditions.

#### T-cell responses to CMV, CLIP, immune activation, and frailty

The accumulation of CMV-specific T-cells as well as extensive T-cell responses to CMV likely contributes to CLIP and immune activation as these cells produce large amounts of inflammatory mediators such as interferon- $\gamma$ , tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and IL-6. In the WHAS II cohort, the presence of CMV UL123 DNA was associated with elevated IL-6 levels [18]. In another study of community-dwelling older adults, the presence of CMV UL123 DNA in monocytes, but not anti-CMV IgG serology, was associated with elevated levels of neopterin, a marker of monocyte and macrophage activation which has also been associated with frailty in older adults [29, 40].

More recent data from 22 virologically suppressed HIV+ and 20 HIV- men from the MACS show that T-cell responses to 19 CMV ORFs are correlated with serum levels of inflammatory mediators [41]. Although CMV titers were not measured in these participants, all men exhibited positive T-cell responses to CMV. Generally speaking, T-cell responses to CMV were broad with high inter-person variation. Strong correlations between both CD8 and CD4 T-cell responses and inflammatory and immune activation markers in the HIV+ non-frail as well as HIV- frail and non-frail participants. Correlations with cytokines and activated CD4 and CD8 T-cells were positive, while correlations with chemokines were generally negative [41]. Furthermore, in these same individuals, CD4 T-cell IL-2 responses to UL83 or UL123 alone were not correlated with elevated levels of CPR, but the total CD4 T-cell IL-2 response to all 19 CMV ORFs was (Fig. 1). In another study of HIV+ individuals, CMV-specific CD4 T-cells responses were associated with CD8 and CD4 T-cell activation, but not with inflammatory markers [42]. However, only T-cell responses to CMV-pp65 and glycoprotein B (gB) were assessed. Once again, this supports the need to investigate the responses to more than a few antigenic epitopes when assessing responses to CMV infection. While these new data support a role for T-cell responses to CMV in chronic inflammation and immune activation, it is important to note that an age-related increase in levels of inflammatory mediators CPR, IL-6, TNF- $\alpha$  is also observed in CMV-seronegative individuals [43]. Thus, other mechanisms may contribute to CLIP and immune activation in older adults.

It has been hypothesized that CMV infection could contribute to frailty through chronic inflammation or CLIP [44]. In the WHAS I and II cohorts, chronic CMV infection assessed by positive CMV IgG serology has been associated with frailty, the association was particularly strong in participants with high IL-6 levels suggesting a possible link between CMV infection and frailty through increased inflammation [24]. We were also able to show that a high CD4 T-cell IL-2 response to CMV was predictive for the onset of frailty in HIV- non-frail men. However, this was not the case for HIV+ non-frail men [41]. Whether HIV infection modifies T-cell response to CMV and/or its relationship with CLIP in aging MSM or general aging population deserves further investigation.



Percent Responding

Fig. 1 Correlations between cytomegalovirus-induced CD4 IL-2 responses and serum C-reactive protein (CRP). CD4 T-cell IL-2 responses to CMV-pp65 (encoded by) UL83 or IE1 (UL123) alone or together are only poorly correlated with serum CRP while the total

## **Concluding remarks**

Recent data support the hypothesis that T-cell responses to CMV could explain much of CLIP and immune activation in people with treated HIV infection and in older adults. However, it remains challenging to distinguish the direct effect of CMV on CLIP and immune activation from other potential etiologic mechanisms. To this end, the role of CMV reactivation should also be further investigated. Additional investigations including longitudinal and treatment studies are needed. These studies should take frailty status into account. Recently developed novel evaluation tools, especially T-cell responses to a broad range of epitopes and diagnostic methods of chronic CMV infection such as nested PCR and ddPCR should facilitate more accurate assessment of the immunological burden of CMV and its impact on CLIP and immune activation, and possibly its role in the pathogenesis of frailty and other age-related chronic conditions.

Acknowledgements Work presented in this review was supported in part by NIH Grants R21-AG-043874 and R01AI108907 to Dr. Sean X. Leng, U01-AI35042 (MACS) to Dr. Joseph B. Margolick, and Irma

CD4 T-cell IL-2 response to peptide pools encoded by all 19 CMV open reading frames is strongly correlated with elevated serum CRP levels

and Paul Milstein Program for Senior Health, Milstein Medical Asian American Partnership (MMAAP) Foundation (http://www.mmaap f.org) to Dr. Sean X. Leng.

#### **Compliance with ethical standards**

**Conflict of interest** The authors declare they have no conflict of interest.

#### References

- Chen YY, Liu S, Leng SX (2019) Chronic low-grade inflammatory phenotype (CLIP), senescent immune dysregulation, and frailty. Clin Ther. https://doi.org/10.1016/j.clinthera.2019.02.001
- Leng SX, Xue Q-L, Tian J, Walston JD, Fried LP (2007) Inflammation and frailty in older women. J Am Geriatr Soc 55:864–871
- Franceschi C, Campisi J (2014) Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. J Gerontol A Biol Sci Med Sci 69(Suppl 1):S4–S9
- Michaud M, Balardy L, Moulis G, Gaudin C, Peyrot C, Vellas B, Cesari M, Nourhashemi F (2013) Proinflammatory cytokines, aging, and age-related diseases. J Am Med Dir Assoc 14:877–882
- Regidor DL, Detels R, Breen EC, Widney DP, Jacobson LP, Palella F, Rinaldo CR, Bream J, Martínez-Maza O (2011) Effect

of highly active antiretroviral therapy on biomarkers of B-lymphocyte activation and inflammation. AIDS 25:303–314

- Wada NI, Jacobson LP, Margolick JB, Breen EC, Macatangay B, Penugonda S, Martínez-Maza O, Bream J (2015) The effect of HAART-induced HIV suppression on circulating markers of inflammation and immune activation. AIDS 29:463–471
- 7. Deeks SG (2011) HIV infection, inflammation, immunosenescence, and aging. Annu Rev Med 62:141–155
- Margolick JB, Bream JH, Martínez-Maza O, Lopez J, Li X, Phair JP, Koletar S, Jacobson LP (2017) Frailty and circulating markers of inflammation in HIV+ and HIV- men in the multicenter AIDS cohort study. J Acquir Immune Defic Syndr 74:407–417
- Lachmann R, Loenenbach A, Waterboer T, Brenner N, Pawlita M, Michel A, Thamm M, Poethko-Müller C, Wichmann O, Wiese-Posselt M (2018) Cytomegalovirus (CMV) seroprevalence in the adult population of Germany. PLoS One 13:e0200267
- Staras SAS, Dollard SC, Radford KW, Flanders WD, Pass RF, Cannon MJ (2006) Seroprevalence of cytomegalovirus infection in the United States, 1988–1994. Clin Infect Dis 43:1143–1151
- Naeger DM, Martin JN, Sinclair E, Hunt PW, Bangsberg DR, Hecht F, Hsue P, McCune JM, Deeks SG (2010) Cytomegalovirus-specific T cells persist at very high levels during long-term antiretroviral treatment of HIV disease. PLoS One 5:e8886
- Ouyang Q, Wagner WM, Zheng W, Wikby A, Remarque EJ, Pawelec G (2004) Dysfunctional CMV-specific CD8(+) T cells accumulate in the elderly. Exp Gerontol 39:607–613
- Ouyang Q, Wagner WM, Wikby A, Walter S, Aubert G, Dodi AI, Travers P, Pawelec G (2003) Large numbers of dysfunctional CD8+ T lymphocytes bearing receptors for a single dominant CMV epitope in the very old. J Clin Immunol 23:247–257
- Vescovini R, Biasini C, Fagnoni FF, Telera AR, Zanlari L, Pedrazzoni M, Bucci L, Monti D, Medici MC, Chezzi C, Franceschi C, Sansoni P (2007) Massive load of functional effector CD4+ and CD8+ T cells against cytomegalovirus in very old subjects. J Immunol 179:4283–4291
- Khan N, Shariff N, Cobbold M, Bruton R, Ainsworth JA, Sinclair AJ, Nayak L, Moss PA (2002) Cytomegalovirus seropositivity drives the CD8 T cell repertoire toward greater clonality in healthy elderly individuals. J Immunol 169:1984–1992
- Pawelec G, Akbar A, Caruso C, Solana R, Grubeck-Loebenstein B, Wikby A (2005) Human immunosenescence: is it infectious? Immunol Rev 205:257–268
- Leng SX, Qu T, Semba RD, Li H, Yao X, Nilles T, Yang X, Manwani B, Walston JD, Ferrucci L, Fried LP, Margolick JB, Bream J (2011) Relationship between cytomegalovirus (CMV) IgG serology, detectable CMV DNA in peripheral monocytes, and CMV pp65495–503-specific CD8+ T cells in older adults. Age 33:607–614
- Li H, Weng P, Najarro K, Xue Q-L, Semba RD, Margolick JB, Leng S (2014) Chronic CMV infection in older women: longitudinal comparisons of CMV DNA in peripheral monocytes, anti-CMV IgG titers, serum IL-6 levels, and CMV pp65 (NLV)specific CD8(+) T-cell frequencies with 12 year follow-up. Exp Gerontol 54:84–89
- Sylwester AW, Mitchell BL, Edgar JB, Taormina C, Pelte C, Ruchti F, Sleath PR, Grabstein KH, Hosken NA, Kern F, Nelson JA, Picker LJ (2005) Broadly targeted human cytomegalovirusspecific CD4+ and CD8+ T cells dominate the memory compartments of exposed subjects. J Exp Med 202:673–685
- Khan N, Hislop A, Gudgeon N, Cobbold M, Khanna R, Nayak L, Rickinson AL, Moss PA (2004) Herpesvirus-specific CD8 T cell immunity in old age: cytomegalovirus impairs the response to a coresident EBV infection. J Immunol 173:7481–7489
- Furman D, Jojic V, Sharma S, Shen-Orr SS, Angel CJL, Onengut-Gumuscu S, Kidd BA, Maecker HT, Concannon P, Dekker CL, Thomas PG, Davis MM (2015) Cytomegalovirus infection

enhances the immune response to influenza. Sci Transl Med 7:281ra43

- 22. Wang GC, Kao WHL, Murakami P, Xue Q-L, Chiou RB, Detrick B, McDyer JF, Semba RD, Casolaro V, Walston JD, Fried LP (2010) Cytomegalovirus infection and the risk of mortality and frailty in older women: a prospective observational cohort study. Am J Epidemiol 171:1144–1152
- 23. Hadrup SR, Strindhall J, Køllgaard T, Seremet T, Johansson B, Pawelec G, Thor Straten P, Wikby A (2006) Longitudinal studies of clonally expanded CD8 T cells reveal a repertoire shrinkage predicting mortality and an increased number of dysfunctional cytomegalovirus-specific T cells in the very elderly. J Immunol 176:2645–2653
- Schmaltz HN, Fried LP, Xue Q-L, Walston J, Leng SX, Semba RD (2005) Chronic cytomegalovirus infection and inflammation are associated with prevalent frailty in community-dwelling older women. J Am Geriatr Soc 53:747–754
- Aiello AE, Haan MN, Pierce CM, Simanek AM, Liang J (2008) Persistent infection, inflammation, and functional impairment in older Latinos. J Gerontol A Biol Sci Med Sci 63:610–618
- Strandberg TE, Pitkala KH, Tilvis RS (2009) Cytomegalovirus antibody level and mortality among community-dwelling older adults with stable cardiovascular disease. JAMA 301:380–382
- Hunt PW, Martin JN, Sinclair E, Epling L, Teague J, Jacobson MA, Tracy RP, Corey L, Deeks SG (2011) Valganciclovir reduces T cell activation in HIV-infected individuals with incomplete CD4+ T cell recovery on antiretroviral therapy. J Infect Dis 203:1474–1483
- Ding X, Margolick JB, Leng SX (2017) T-cell immunity against cytomegalovirus in older adults. In: Fulop T, Franceschi C, Hirokawa K, Pawelec G (eds) Handbook of immunosenescence: basic understanding and clinical implications. Springer International Publishing, Cham, pp 1–14
- Leng SX, Li H, Xue Q-L, Tian J, Yang X, Ferrucci L, Fedarko N, Fried LP, Semba RD (2011) Association of detectable cytomegalovirus (CMV) DNA in monocytes rather than positive CMV IgG serology with elevated neopterin levels in community-dwelling older adults. Exp Gerontol 46:679–684
- Matheï C, Adriaensen W, Vaes B, Van Pottelbergh G, Wallemacq P, Degryse J (2015) No relation between CMV infection and mortality in the oldest old: results from the Belfrail study. Age Ageing 44:130–135
- Parry HM, Zuo J, Frumento G, Mirajkar N, Inman C, Edwards E, Griffiths M, Pratt G, Moss P (2016) Cytomegalovirus viral load within blood increases markedly in healthy people over the age of 70 years. Immun Ageing 13:1
- Lachmann R, Bajwa M, Vita S, Smith H, Cheek E, Akbar A, Kern F (2012) Polyfunctional T cells accumulate in large human cytomegalovirus-specific T cell responses. J Virol 86:1001–1009
- Moss P, Khan N (2004) CD8(+) T-cell immunity to cytomegalovirus. Hum Immunol 65:456–464
- Pita-Lopez ML, Gayoso I, DelaRosa O, Casado JG, Alonso C, Muñoz-Gomariz E, Tarazona R, Solana R (2009) Effect of ageing on CMV-specific CD8 T cells from CMV seropositive healthy donors. Immun Ageing 6:11
- 35. Kern F, Faulhaber N, Frömmel C, Khatamzas E, Prösch S, Schönemann C, Kretzschmar I, Volkmer-Engert R, Volk HD, Reinke P (2000) Analysis of CD8 T cell reactivity to cytomegalovirus using protein-spanning pools of overlapping pentadecapeptides. Eur J Immunol 30:1676–1682
- Stern-Ginossar N, Weisburd B, Michalski A, Khanh Le VT, Hein MY, Huang S-X, Ma M, Shen B, Qian S, Hengel H, Mann M, Ingolia NT, Weissman JS (2012) Decoding human cytomegalovirus. Science 338:1088–1093
- Li H, Margolick JB, Bream JH, Nilles TL, Langan S, Bui HT, Sylwester AW, Picker LJ, Leng SX (2014) Heterogeneity of CD4+

and CD8+ T-cell responses to cytomegalovirus in HIV-infected and HIV-uninfected men who have sex with men. J Infect Dis 210:400-404

- Jackson SE, Mason GM, Okecha G, Sissons JGP, Wills MR (2014) Diverse specificities, phenotypes, and antiviral activities of cytomegalovirus-specific CD8+ T cells. J Virol 88:10894–10908
- 39. Bajwa M, Vita S, Vescovini R, Larsen M, Sansoni P, Terrazzini N, Caserta S, Thomas D, Davies KA, Smith H. Kern F (2016) Functional diversity of cytomegalovirus-specific T cells is maintained in older people and significantly associated with protein specificity and response size. J Infect Dis 214:1430–1437
- Leng SX, Tian X, Matteini A, Li H, Hughes J, Jain A, Walston JD, Fedarko NS (2011) IL-6-independent association of elevated serum neopterin levels with prevalent frailty in community-dwelling older adults. Age Ageing 40:475–481
- 41. Margolick JB, Bream JH, Nilles TL, Li H, Langan SJ, Deng S, Wang R, Wada N, Leng SX (2018) Relationship between T-cell responses to CMV, markers of inflammation, and frailty in HIVuninfected and HIV-infected men in the multicenter AIDS cohort study. J Infect Dis 218:249–258

- 42. Ballegaard V, Brændstrup P, Pedersen KK, Kirkby N, Stryhn A, Ryder LP, Gerstoft J, Nielsen SD (2018) Cytomegalovirus-specific T-cells are associated with immune senescence, but not with systemic inflammation, in people living with HIV. Sci Rep 8:3778
- 43. Bartlett DB, Firth CM, Phillips AC, Moss P, Baylis D, Syddall H, Sayer AA, Cooper C, Lord JM (2012) The age-related increase in low-grade systemic inflammation (Inflammaging) is not driven by cytomegalovirus infection. Aging Cell 11:912–915
- Wang GC, Walston J (2009) CMV infection and frailty: immunologic consequences and disease pathogenesis. In: Fulop T, Franceschi C, Hirokawa K, Pawelec G (eds) Handbook on immunosenescence: basic understanding and clinical applications. Springer, Berlin, pp 1305–1326

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.